

Research Article

Species diversity and taxonomy of *Vararia* (Russulales, Basidiomycota) with descriptions of six species from Southwestern China

Yinglian Deng^{1,2}, Sana Jabeen³, Changlin Zhao^{1,2,4}

- The Key Laboratory of Forest Resources Conservation and Utilization in the South-west Mountains of China Ministry of Education, Key Laboratory of National Forestry and Grassland Administration on Biodiversity Conservation in Southwest China, Yunnan Provincial Key Laboratory for Conservation and Utilization of In-forest Re-source, Southwest Forestry University, Kunming 650224, China
- 2 College of Biodiversity Conservation, Southwest Forestry University, Kunming 650224, China
- 3 Department of Botany, Division of Science and Technology, University of Education, Township, Lahore, Punjab, Pakistan
- 4 Yunnan Academy of Biodiversity, Southwest Forestry University, Kunming 650224, China

Corresponding author: Changlin Zhao (fungi@swfu.edu.cn)

Abstract

Vararia is a species-rich genus in the family Peniophoraceae and has been shown to be polyphyletic. In this study, sequences of ITS and LSU rRNA markers of the studied samples were generated and phylogenetic analyses were performed with the maximum likelihood, maximum parsimony, and Bayesian inference methods. Seventeen lineages including six new species from China, i.e., V. fissurata, V. lincangensis, V. punctata, V. isabellina, V. sinensis, and V. yaoshanensis were recognized, in which V. fissurata is characterized by the brittle basidiomata with pruinose and cracking hymenophore having white to olivaceous buff hymenial surface, the clamped generative hyphae, presence of the two types gloeocystidia; V. lincangensis is characterized by the simple-septa generative hyphae, and thick-walled skeletal hyphae, and ellipsoid basidiospores; V. punctata is delimited by its thin to slightly thick-walled generative hyphae, and thick-walled skeletal hyphae, present thick-walled, clavate to cylindrical gloeocystidia; V. isabellina is characterized by having the cream to isabelline to slightly brown hymenial surface, thin to slightly thick-walled generative hyphae, and sub-fusiform to navicular basidiospores; V. sinensis is distinguishable by its white to slightly pink hymenial surface, thick-walled skeletal hyphae, and sub-fusiform to navicular basidiospores; V. yaoshanensis is characterized by cream to pinkish buff to cinnamon-buff hymenial surface, slightly thick-walled generative hyphae, the presence of two types gloeocystidia, and slightly thick-walled, ellipsoid basidiospores. Phylogram based on the ITS+nLSU rDNA gene regions included nine genera within the family Peniophoraceae as Amylostereum, Asterostroma, Baltazaria, Dichostereum, Michenera, Peniophora, Scytinostroma and Vararia, in which the six new wood-inhabiting fungi species were grouped into genus Vararia. The phylogenetic tree inferred from the combined ITS and LSU tree sequences highlighted that V. fissurata was found to be the sister to V. ellipsospora with strong supports. Additionally, V. lincangensis was clustered with V. fragilis. Furthermore, V. punctata was retrieved as a sister to V. ambigua. Moreover, V. sinensis was grouped with five taxa as V. breviphysa, V. pirispora, V. fusispora, V. abortiphysa and V. insolita. The new species V. isabellina formed a monophyletic lineage, in which it was then grouped closely with V. daweishanensis, and V. gracilispora. In addition, V. yaoshanensis was found to be the sister to V. gallica with strong supports. The present results increased the knowledge of Vararia species diversity and taxonomy of corticioid fungi in China. An identification key to 17 species of *Vararia* in China is provided.



Academic editor: María P. Martín Received: 17 January 2024 Accepted: 1 March 2024 Published: 22 March 2024

Citation: Deng Y, Jabeen S, Zhao C (2024) Species diversity and taxonomy of *Vararia* (Russulales, Basidiomycota) with descriptions of six species from Southwestern China. MycoKeys 103: 97–128. https://doi.org/10.3897/mycokeys.103.118980

Copyright: © Yinglian Deng et al.
This is an open access article distributed under terms of the Creative Commons Attribution
License (Attribution 4.0 International – CC BY 4.0).

Key words: Biodiversity, China, phylogenetic analyses, taxonomy, wood-inhabiting fungi, Yunnan Province

Introduction

Fungi represent one of the most diverse groups of organisms on the earth, with an indispensable role in the processes and functioning of forest ecosystems (Hyde 2022). The genus *Vararia* P. Karst. belongs to the family Peniophoraceae of the order Russulales (Larsson and Larsson 2003; Miller et al. 2006; Larsson 2007). The Russulales is a well-known order that contains morphologically diverse mushrooms (Miller et al. 2006). Species from this order comprise many representative wood-inhabiting fungal taxa, including hydnoid, corticioid, and polyporoid basidiomes with diverse hymenophoral and cystidial morphology (Yurchenko and Wu 2016; Riebesehl and Langer 2017; Yurchenko et al. 2017; Cui et al. 2019; Riebesehl et al. 2019; Jiang et al. 2021; Wu et al. 2022).

The genus Vararia is a corticioid wood-inhabiting fungal genus with a wide distribution, typified by V. investiens (Schwein.) P. Karst. It was first described by Karsten as a subgenus of Xerocarpus P. Karst. for Xerocarpus alutarius (Berk. & M. A. Curtis) P. Karst., which was later found to be a synonym of Radulum investiens Schwein. Karsten raised Xerocarpus subgen. Vararia to the generic rank (Karasinski 2010). The genus is characterized by the resupinate basidiomata, a dimitic hyphal structure with simple-septate or clamped generative hyphae and often dextrinoid dichohyphae in Melzer's reagent, the presence of gloeocystidia, and variously shaped smooth basidiospores with or without an amyloid reaction (Karnste 1898; Boidin and Lanquetin 1975; Boidin 1980; Bernicchia and Gorjón 2010). The species of Vararia are found on fallen angiosperm branches, dead woody or herbaceous stems or occasionally on gymnosperm wood (Yurchenko et al. 2017). Based on the MycoBank database (http://www.mycobank.org, accessed on 17 January 2024) and the Index Fungorum (http://www.indexfungorum.org, accessed on 17 January 2024), Vararia has registered 99 specific and infraspecific names, and the actual number of the species has reached up to 76, currently known, and they occur mainly in the tropical and subtropical areas of the world (Cunningham 1955; Gilbertson 1965; Boidin 1967; Pouzar 1982; Boidin and Languetin 1987; Stalpers 1996; Boidin and Gilles 1999; Larsson and Larsson 2003; Bernicchia and Gorjón 2010; Duhem and Buyck 2012; Sanyal et al. 2012; Nakasone 2015; Liu and He 2016; Dai et al. 2021; Zou et al. 2022; Deng and Zhao 2023).

Classification of the kingdom of fungi has been updated continuously, based on the frequent inclusion of data from DNA sequences in many phylogenetic studies (Yurchenko et al. 2020). These pioneering research studies into the family Peniophoraceae were just the prelude to the molecular systematics period (Zou et al. 2022). The phylogenetic diversity displayed by corticioid fungal species, based on ITS1-5.8S-ITS2-nrLSU nuclear rDNA, revealed that the taxa of Peniophoraceae were nested in the russuloid clade, which holds a considerable share of the phylogenetic framework, and included the genera of Asterostroma Massee, Baltazaria Leal-Dutra, Dentinger & G.W. Griff., Dichostereum Pilát, Gloiothele Bres., Lachnocladium Lév., Michenera Berk. & M.A. Curtis, Peniophora Cooke, Scytinostroma Donk, Vesiculomyces E. Hagstr. and Vararia (Larsson and

Larsson 2003; Larsson and Larsson 2004; Larsson 2007; Leal-Dutra et al. 2018; Zou et al. 2022; Li et al. 2023). Morphologically, *Scytinostroma* was similar to *Vararia*, which usually differed in having the typical dichohyphae (Bernicchia and Gorjón 2010). The taxonomic distinction between *Scytinostroma* and *Vararia* has been questioned (Hallenberg 1985; Boidin and Lanquetin 1987; Stalpers 1996; Boidin et al 1998). However, there has been general agreement that the two genera were closely related and that they together made up a natural group. Larsson and Larsson (2003) strongly suggested that neither skeletal hyphae nor their branching patterns have any predictive power in a phylogenetic context.

During investigations on the wood-inhabiting fungi in the Yunnan province of China, the samples representing six additional species belonging to genera *Vararia* were collected. To clarify the placement and relationships of the six species, we carried out a phylogenetic and taxonomic study on *Vararia*, based on the ITS and LSU sequences.

Materials and methods

Morphology

Fresh fruiting bodies of the fungi were collected from Dali, Dehong, Lincang, Puer, Yuxi and Zhaotong of Yunnan Province, P.R. China. Specimens were dried in an electric food dehydrator at 40 °C, then sealed and stored in an envelope bag and deposited in the herbarium of the Southwest Forestry University (SWFC), Kunming, Yunnan Province, P.R. China. Macromorphological descriptions are based on field notes and photos captured in the field and lab. Color terminology follows Petersen (Petersen 1996). Micromorphological data were obtained from the dried specimens when observed under a light microscope following the previous study (Zhao et al. 2023; Guan et al. 2023). The following abbreviations are used: KOH = 5% potassium hydroxide water solution, CB = Cotton Blue, CB = acyanophilous, IKI = Melzer's Reagent, IKI = both inamyloid and indextrinoid, L = mean spore length (arithmetic average for all spores), W = mean spore width (arithmetic average for all spores), Q = variation in the L/W ratios between the specimens studied and n = a/b (number of spores (a) measured from given number (b) of specimens).

Molecular phylogeny

The EZNA HP Fungal DNA Kit (Omega Biotechnologies Co., Ltd., Kunming, China) was used to extract DNA with some modifications from the dried specimens. The nuclear ribosomal ITS region was amplified with primers ITS5 and ITS4 (White et al. 1990). The PCR procedure for ITS was as follows: initial denaturation at 95 °C for 3 min, followed by 35 cycles at 94 °C for 40 s, 58 °C for 45 s and 72 °C for 1 min, and a final extension of 72 °C for 10 min. The nuclear LSU region was amplified with primer pair LROR and LR7 (Vilgalys and Hester 1990; Rehner and Samuels 1994). The PCR procedure for LSU was as follows: initial denaturation at 94 °C for 1 min, followed by 35 cycles at 94 °C for 30 s, 48 °C for 1 min and 72 °C for 1.5 min, and a final extension of 72 °C for 10 min. The PCR procedure for ITS and LSU followed a previous study (Zhao and Wu 2017). All of the newly generated sequences were deposited in NCBI GenBank (https://www.ncbi.nlm.nih.gov/genbank/) (Table 1).

Table 1. List of species, specimens and GenBank accession numbers of sequences used in this study. [* Indicates type materials].

Species name	Specimen No.	GenBank accession No.		Country	References
		ITS	nLSU		
Amylostereum chailletii	NH8031	AF506406	AF506406	Sweden	Larsson and Larsson 2003
A. laevigatum	NH12863	AF506407	AF506407	Sweden	Larsson and Larsson 2003
Asterostroma bambusicola	He4132	KY263865	KY263871	Thailand	Liu et al. 2017
A. cervicolor	He2314	KY263859	KY263869	China	Unpublished
A. cervicolor	He4020	KY263860	KY263868	Thailand	Unpublished
A. muscicola	He4397	MK625630	MK625563	China	Unpublished
Baltazaria galactina	He4999	MK625618	MK625547	China	Unpublished
B. octopodites	FLOR63715	MH260042	MH260060	United Kingdom	Leal-Dutra et al. 2018
Confertobasidium olivaceoalbum	FP90196	AF511648	AF511648	Sweden	Larsson and Larsson 2003
Dichostereum boidinii	He4410	MH538315	MH538331	China	Vu et al. 2019
D. boidinii	He5026	MH538324	MH538330	China	Liu et al. 2019
D. pallescens	CBS:718.81	MH861456	MH873198	USA	Vu et al. 2019
Metulodontia nivea	NH13108	AF506423	AF506423	Sweden	Larsson and Larsson 2003
Michenera artocreas	GHL-2016-Oct	MH204688	MH204691	USA	Liu et al. 2019
M. incrustata	He5368	MH204689	MH204690	China	Liu et al. 2019
Peniophora cinerea	CBS:261.37	MH855905	MH867412	Belgium	Vu et al. 2019
P. cinerea	He3725	MK588769	MK588809	China	Unpublished
P. incarnata	CBS 430.72	MH860518	MH872230	Netherlands	Vu et al. 2019
P. incarnata	NH10271	AF506425	AF506425	Sweden	Larsson and Larsson 2003
P. nuda	LZ15-07	MT859929	_	China	Unpublished
P. quercina	CBS 407.50	MH856687	MH868204	France	Vu et al. 2019
P. quercina	CBS:410.50	MH856690	MH868207	France	Vu et al. 2019
Scytinostroma acystidiatum	He5646	MK625568	MK625494	China	Unpublished
S. alutum	CBS:762.81	MH861482	MH873221	France	Vu et al. 2019
S. beijingensis	He7768	OQ731943	OQ729731	China	Li et al. 2023
S. boidinii	He6911	OQ731934	OQ729724	China	Li et al. 2023
S. duriusculum	He3590	MK625571	MK625499	China	Unpublished
S. hemidichophyticum	CBS:702.84	MH861818	MH873509	Belgium	Vu et al. 2019
S. renisporum	CBS:771.86	MH862051	MH873738	Bali	Vu et al. 2019
S. subrenisporum	He4792	MK625566	MK625493	China	Unpublished
Vararia abortiphysa	CBS:632.81	MH861387	_	Gabon	Vu et al. 2019
V. ambigua	CBS 634.81	MH861388	MH873137	France	Vu et al. 2019
V. amphithallica	CBS:635.81	MH861389	MH873138	Gabon	Vu et al. 2019
V. amphithallica	CBS:687.81	MH861431	_	France	Vu et al. 2019
V.aurantiaca	CBS:641.81	MH861393	_	France	Vu et al. 2019
V. aurantiaca	CBS:642.81	MH861394	_	Gabon	Vu et al. 2019
V. breviphysa	CBS:643.81	MH873144	MH873144	Gabon	Vu et al. 2019
V. breviphysa	CBS:644.81	MH861396	_	Gabon	Vu et al. 2019
V. calami	CBS:646.81	MH861398	_	France	Vu et al. 2019
V. calami	CBS:648.81	MH861399	_	France	Vu et al. 2019
V. callichroa	CBS:744.91	MH874000	MH874000	France	Vu et al. 2019

Species name	Specimen No.	GenBank accession No.		Country	References
		ITS	nLSU	-	
V. cinnamomea	CBS:641.84	MH861794	_	Madagascar	Vu et al. 2019
V. cinnamomea	CBS:642.84	MH873488	MH873488	Madagascar	Vu et al. 2019
V. cremea	CBS:651.81	MH873147	MH873147	France	Vu et al. 2019
V. daweishanensis	CLZhao 17911	OP380613	_	China	Zou et al. 2022
V. daweishanensis	CLZhao 17936	OP380614	_	China	Zou et al. 2022
V. dussii	CBS:652.81	MH873148	MH873148	France	Vu et al. 2019
V. dussii	CBS:655.81	MH861405	_	France	Vu et al. 2019
V. ellipsospora	HHB-19503	MW740328	_	New Zealand	Zou et al. 2022
V. fissurata	CLZhao 10118	PP083288	_	China	Present study
V. fissurata	CLZhao 10181	PP083289	_	China	Present study
V. fissurata	CLZhao 22538	PP083290	_	China	Present study
V. fissurata	CLZhao 4614	PP083283	_	China	Present study
V. fissurata	CLZhao 5218	OQ025218	OR539502	China	Present study
V. fissurata	CLZhao 6070	PP083284	_	China	Present study
V. fissurata	CLZhao 8171*	OQ025219	OR539503	China	Present study
V. fissurata	CLZhao 9618	PP083285	_	China	Present study
V. fissurata	CLZhao 9668	PP083286	_	China	Present study
V. fissurata	CLZhao 9697	PP083287	_	China	Present study
V. fragilis	CLZhao 16475	OP380612	_	China	Zou et al. 2022
V. fragilis	CLZhao 2628	OP380611	_	China	Zou et al. 2022
V. fusispora	PDD:119539	OL709443	_	New Zealand	Zou et al. 2022
V. gallica	CBS 234.91	MH862250	MH873932	Canada	Vu et al. 2019
V. gallica	CBS 656.81	MH861406	MH873152	France	Vu et al. 2019
V. gillesii	CBS:660.81	MH873153	MH873153	Cote d'Ivoire	Vu et al. 2019
V. gomezii	CBS:661.81	MH873154	MH873154	France	Vu et al. 2019
V. gracilispora	CBS:663.81	MH861411	_	Gabon	Vu et al. 2019
V. gracilispora	CBS:664.81	MH861412	_	Gabon	Vu et al. 2019
V. insolita	CBS:668.81	MH861413	_	France	Vu et al. 2019
V. intricata	CBS:673.81	MH861418	_	France	Vu et al. 2019
V. investiens	FP-151122ITS	MH971976	_	USA	Liu et al. 2019
V. investiens	UC2023140	KP814286	_	USA	Rosenthal et al. 2017
V. isabellina	CLZhao 22852	OR048789	OR506350	China	Present study
V. isabellina	CLZhao 22887	OR048788	OR506351	China	Present study
V. lincangensis	CLZhao 22791*	OR048819	OR506348	China	Present study
V. lincangensis	CLZhao 22799	OR048818	OR506349	China	Present study
V. malaysiana	CBS:644.84	MH873490	MH873490	Singapore	Vu et al. 2019
V. minispora	CBS:682.81	MH861426	_	France	Vu et al. 2019
V. ochroleuca	CBS:465.61	MH858109	_	France	Vu et al. 2019
V. ochroleuca	JS24400	AF506485	AF506485	Norway	Larsson and Larsson 2003
V. parmastoi	CBS:879.84	MH861852	MH861852	Uzbekistan	Vu et al. 2019
V. pectinata	CBS:685.81	MH861429	_	Cote d'Ivoire	Vu et al. 2019
V. perplexa	CBS:695.81	MH861438	_	France	Vu et al. 2019
V. pirispora	CBS:720.86	MH862016	_	France	Vu et al. 2019
V. punctata	CLZhao 22423	OR048813	OR539685	China	Present study

Species name	Specimen No.	GenBank ac	cession No.	Country	References
		ITS	nLSU		
V. punctata	CLZhao 22439*	OR048812	OR510675	China	Present study
V. rhombospora	CBS:743.81	MH861470	_	France	Vu et al. 2019
V. rosulenta	CBS:743.86	MH862028	_	France	Vu et al. 2019
V. rugosispora	CBS:697.81	MH861440	_	Gabon	Vu et al. 2019
V. sigmatospora	CBS:748.91	MH874001	MH874001	Netherlands	Vu et al. 2019
V. sinensis	CLZhao 25160*	OR102494	OR510678	China	Present study
V. sinensis	CLZhao 25161	OR102495	OR510679	China	Present study
V. sphaericospora	CBS:700.81	MH873185	MH873185	Gabon	Vu et al. 2019
V. sphaericospora	CBS:703.81	MH861446	_	Gabon	Vu et al. 2019
V. sphaericospora	He4847	MK625592	MK625521	China	Unpublished
V. trinidadensis	CBS:650.84	MH873495	MH873495	Madagascar	Vu et al. 2019
V. trinidadensis	CBS:651.84	MH861803	_	Madagascar	Vu et al. 2019
V. tropica	CBS 704.81	MH861447	MH873189	France	Vu et al. 2019
V. vassilievae	UC2022892	KP814203	_	USA	Unpublished
V. verrucosa	CBS:706.81	MH861449	MH861449	France	Vu et al. 2019
V. yaoshanensis	CLZhao 20528	PP091673	_	China	Present study
V. yaoshanensis	CLZhao 20531	PP091674	_	China	Present study
V. yaoshanensis	CLZhao 20565	PP091675	PP091683	China	Present study
V. yaoshanensis	CLZhao 20605	PP091676	_	China	Present study
V. yaoshanensis	CLZhao 20608	PP091677	_	China	Present study
V. yaoshanensis	CLZhao 20617	PP091678	_	China	Present study
V. yaoshanensis	CLZhao 20619	PP091679	_	China	Present study
V. yaoshanensis	CLZhao 20624	PP091680	_	China	Present study
V. yaoshanensis	CLZhao 20646	PP091681	_	China	Present study
V. yaoshanensis	CLZhao 20656	PP091682	_	China	Present study
V. yaoshanensis	CLZhao 20669	PP091666	_	China	Present study
V. yaoshanensis	CLZhao 20677	PP091667	_	China	Present study
V. yaoshanensis	CLZhao 20693*	PP091665	PP091684	China	Present study
V. yaoshanensis	CLZhao 20697	PP091668	_	China	Present study
V. yaoshanensis	CLZhao 20709	PP091669	_	China	Present study
V. yaoshanensis	CLZhao 20713	PP091670	_	China	Present study
V. yaoshanensis	CLZhao 20717	PP091671	_	China	Present study
V. yaoshanensis	CLZhao 20724	PP091672	_	China	Present study

The sequences were aligned in MAFFT version 7 (Katoh et al. 2019) using the G-INS-i strategy. The alignment was adjusted manually using AliView version 1.27 (Larsson 2014). Sequences of *Confertobasidium olivaceoalbum* (Bourdot & Galzin) (AF511648) Jülich and *Metulodontia nivea* (P. Karst.) Parmasto () retrieved from GenBank were used as the outgroups in the ITS+LSU analysis (Fig. 1); Sequences of *Peniophora incarnata* (Pers.) P. Karst. (AF506425) and *Peniophora nuda* (Fr.) Bres. (MT859929) retrieved from GenBank were used as the outgroups in the ITS analysis (Fig. 2) (Leal-Dutra et al. 2018; Zhao et al. 2021).

Maximum parsimony (MP), Maximum Likelihood (ML), and Bayesian Inference (BI) analyses were applied to the combined three datasets following

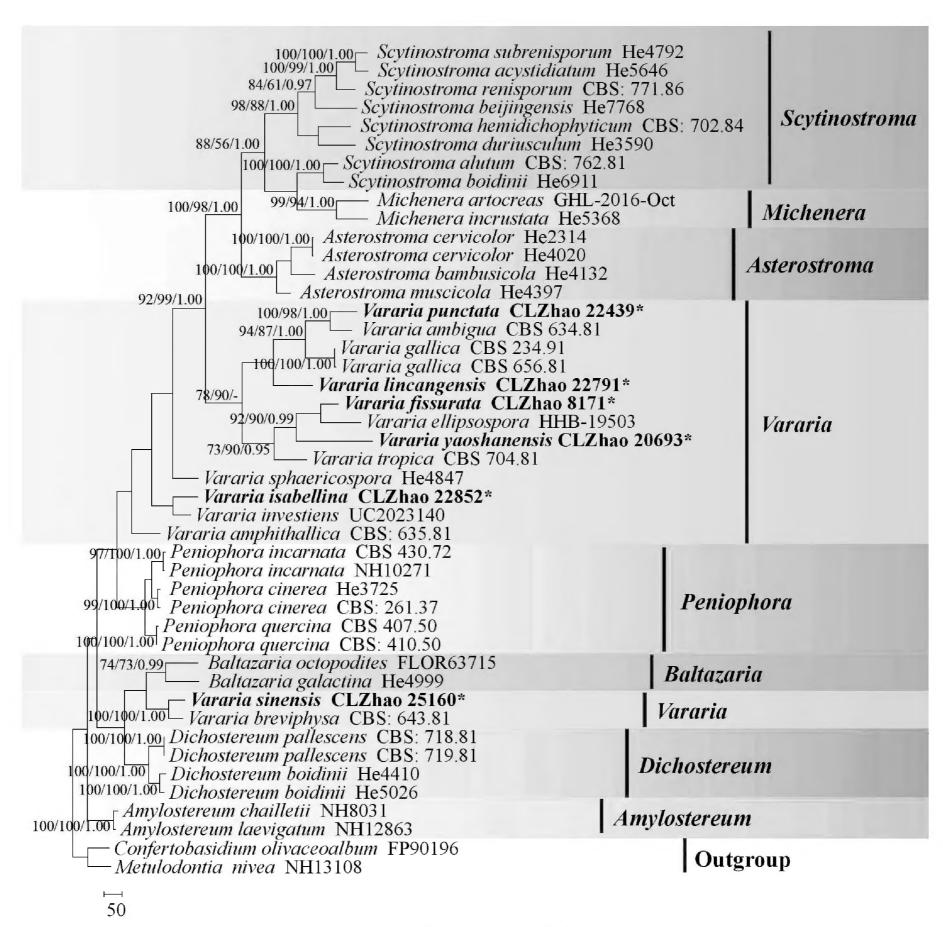


Figure 1. Maximum parsimony strict consensus tree illustrating the phylogeny of *Vararia* and related genera in the family Peniophoraceae based on ITS+LSU sequences. Branches are labelled with maximum likelihood bootstrap values > 70%, parsimony bootstrap values > 50% and Bayesian posterior probabilities > 0.95, respectively.

a previous study (Zhao and Wu 2017). All characters were equally weighted and gaps were treated as missing data. Trees were inferred using the heuristic search option with TBR branch swapping and 1,000 random sequence additions. Max-trees were set to 5,000, branches of zero length were collapsed and all parsimonious trees were saved. Clade robustness was assessed using bootstrap (BT) analysis with 1,000 pseudo replicates (Felsenstein 1985). Descriptive tree statistics – tree length (TL), composite consistency index (Cl), composite retention index (RI), composite rescaled consistency index (RC) and composite homoplasy index (HI) – were calculated for each maximum parsimonious tree generated. The combined dataset was also analysed using Maximum Likelihood (ML) in RAxML-HPC2 through the CIPRES Science Gateway (Miller et al. 2012). Branch support (BS) for the ML analysis was determined by 1000 bootstrap pseudo replicates.

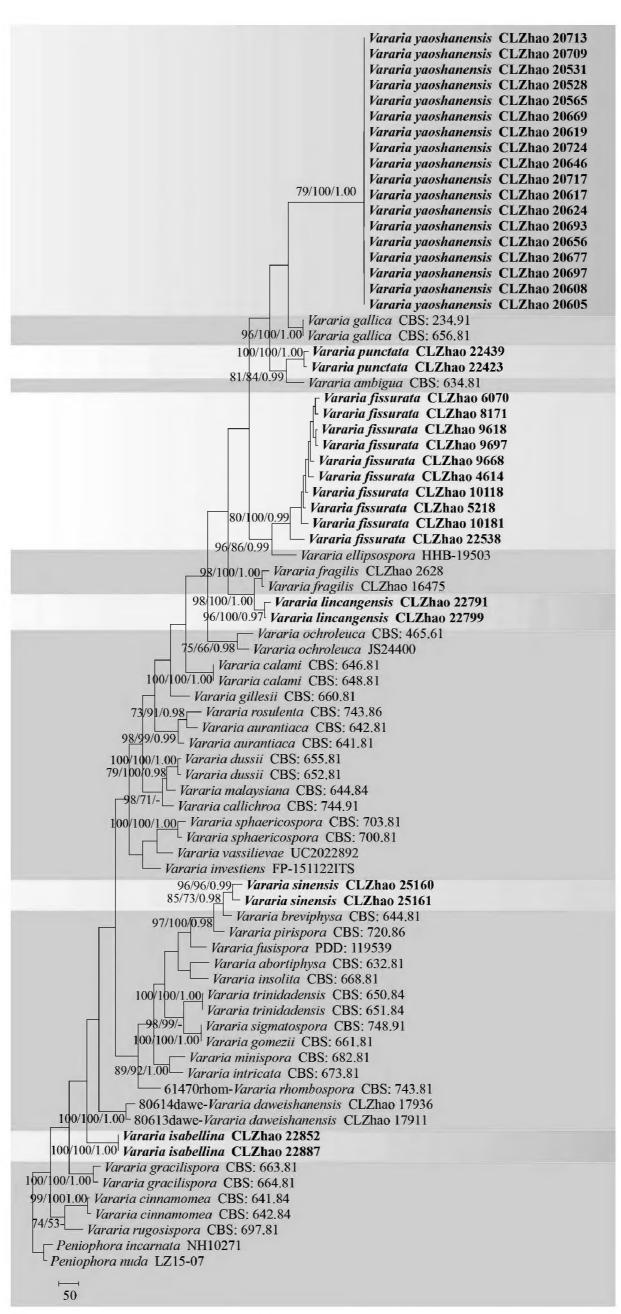


Figure 2. Maximum parsimony strict consensus tree illustrating the phylogeny of the two new species and related species in *Vararia*, based on ITS sequences. Branches are labelled with maximum likelihood bootstrap values > 70%, parsimony bootstrap values > 50% and Bayesian posterior probabilities > 0.95, respectively.

MrModeltest 2.3 (Nylander 2004) was used to determine the best-fit evolution model for each dataset for the purposes of Bayesian inference (BI) which was performed using MrBayes 3.2.7a with a GTR+I+G model of DNA substitution and a gamma distribution rate variation across sites (Ronquist et al. 2012). A total of four Markov chains were run for two runs from random starting trees for 1.2 million generations for ITS+LSU (Fig. 1); and 4 million generations for ITS (Fig. 2) with trees and parameters sampled every 1,000 generations. The first quarter of all the generations were discarded as burn-ins. A majority rule consensus tree was computed from the remaining trees. Branches were considered as significantly supported if they received a maximum likelihood bootstrap support value (BS) of > 70%, a maximum parsimony bootstrap support value (BT) of > 70% or a Bayesian posterior probability (BPP) of > 0.95.

Results

Molecular phylogeny

The ITS+LSU dataset (Fig. 1) comprised sequences from 45 fungal specimens representing 38 taxa. The dataset had an aligned length of 2,304 characters, of which 1,181 characters were constant, 346 were variable and parsimony-uninformative and 777 (50%) were parsimony-informative. Maximum parsimony analysis yielded 3 equally parsimonious trees (TL = 5,051, CI = 0.3985, HI = 0.6015, RI = 0.5522 and RC = 0.2201). The best model of nucleotide evolution for the ITS+LSU dataset estimated and applied in the Bayesian analysis was found to be GTR+I+G. Bayesian analysis and ML analysis resulted in a similar topology as in the MP analysis. The Bayesian analysis had an average standard deviation of split frequencies = 0.004451 (BI) and the effective sample size (ESS) across the two runs is double the average ESS (avg. ESS) = 324. The phylogram based on the ITS+LSU rDNA gene regions (Fig. 1) included eight genera within Peniophoraceae (Russulales), which were Asterostroma, Amylostereum, Baltazaria, Dichostereum, Michenera, Peniophora, Scytinostroma and Vararia, in which six new species were grouped into the genera Vararia.

The ITS dataset (Fig. 2) comprised sequences from 79 fungal specimens representing 38 taxa. The dataset had an aligned length of 849 characters, of which 199 characters were constant, 65 were variable and parsimony-uninformative and 585 (50%) were parsimony-informative. Maximum parsimony analysis yielded 1 equally parsimonious tree (TL = 4,058, CI = 0.3233, HI = 0.6767, RI = 0.7299 and RC = 0.2360). The best model of nucleotide evolution for the ITS dataset estimated and applied in the Bayesian analysis was found to be GTR+I+G. Bayesian analysis and ML analysis resulted in a similar topology as in the MP analysis. The Bayesian analysis had an average standard deviation of split frequencies = 0.001947 (BI) and the effective sample size (ESS) across the two runs is double the average ESS (avg. ESS) = 888. The phylogenetic tree (Fig. 2), inferred from the ITS sequences, highlighted that V. fissurata was the sister to *V. ellipsospora* G. Cunn. with strong supports. The new species V. lincangensis was clustered with V. fragilis L. Zou & C.L. Zhao. Furthermore, V. punctata was retrieved as a sister to V. ambigua Boidin, Lanq. & Gilles. Moreover, V. isabellina formed a monophyletic lineage, and it was then grouped closely with V. daweishanensis L. Zou & C.L. Zhao, and V. gracilispora Boidin & Lanq. The species *V. sinensis* was grouped with five taxa as *Vararia breviphysa* Boidin & Lanq., *V. pirispora* Boidin, Gilles & Lanq., *V. fusispora* G. Cunn., *V. abortiphysa* Boidin & Lanq., and *V. insolita* Boidin & Lanq. In addition, *V. yaoshanensis* was sister to *V. gallica* (Bourdot & Galzin) Boidin with strong supports.

Taxonomy

Vararia fissurata Y.L. Deng & C.L. Zhao, sp. nov.

MycoBank No: MB851793

Figs 3, 4

Holotype. China. Yunnan Province, Yuxi, Xinping County, the Ancient Tea Horse Road, 23°57′10″N, 101°30′41″E, altitude 2600 m a.s.l., on the trunk of angiosperm, leg. C.L. Zhao, 21 August 2018, CLZhao 8171 (SWFC).

Etymology. Fissurata (Lat.): referring to the cracking hymenial surface.

Description. Basidiomata annual, resupinate, adnate, pruinose, brittle, without odor or taste when fresh, up to 12 cm long, 2.5 cm wide, and 100 μ m thick. Hymenial surface smooth, white to olivaceous buff when fresh, and olivaceous buff upon drying, sparsely and deeply cracked with age. Sterile margin distinct, white, and up to 2 mm wide.

Hyphal system dimitic, generative hyphae with clamp connections, colorless, thin-walled, moderately branched, interwoven, $2-3~\mu m$ in diameter; IKI–, CB–, tissues unchanged in KOH. Dichohyphae predominate, yellowish, capillary, frequently branched, 1.5 μm in diameter, thick-walled, dichotomously to irregularly branched with main branches and acute tips, weakly to moderately dextrinoid in Melzer's reagent, CB–, tissues unchanged in KOH; subhymenial hyphae densely covered by a lot of bulk crystals.

Gloeocystidia empty or filled with refractive flocculent matter, two types: (1) Gloeocystidia subglobose, colorless, thin-walled, smooth, $11-23 \times 6-12 \mu m$; (2) Gloeocystidia subulate, usually containing refractive materials; slightly constricted at the neck, colorless, thin-walled, smooth, $25.5-43 \times 7-11 \mu m$. Basidia cylindrical, with four sterigmata and a basal clamp connection, $20-27 \times 4-8 \mu m$; basidioles dominant, in shape similar to basidia but slightly smaller.

Basidiospores ellipsoid to broadly ellipsoid, colorless, thin-walled, smooth, IKI-, CB-, $5-10 \times 3-7 \mu m$, L = $7.37 \mu m$, W = $5.22 \mu m$, Q = 1.38-1.44 (n = 150/5).

Additional specimens examined (paratypes). CHINA. Yunnan Province, Yuxi, Xinping County, the Ancient Tea Horse Road, 23°57'10"N, 101°30'41"E, altitude 2600 m a.s.l., on fallen angiosperm branch, leg. C.L. Zhao, 13 January 2018, CLZ-hao 5218 (SWFC); Puer, Zhenyuan County, Heping Town, Damoshan, 23°56'21"N, 101°25'32"E, altitude 2240 m a.s.l., on fallen angiosperm branch, leg. C.L. Zhao, 16 January 2018, CLZhao 6070 (SWFC); Dali, Weishan Country, Qinghua Town, Green Peacock Nature Reserve, 25°23'35"N, 100°31'39"E, altitude 1500 m a.s.l., on the fallen branch of angiosperm, leg. C.L. Zhao, 18 July 2022, CLZhao 22538 (SWFC); Puer, Jingdong County, Wuliangshan National Nature Reserve, 24°34'45"N, 100°830'03"E, altitude 2000 m a.s.l., on fallen angiosperm branch, leg. C.L. Zhao, 6 October 2017, CLZhao 4614 (SWFC); 6 January 2019, CLZhao 9618, CLZhao 9668 and CLZhao 9697 (SWFC); Dali, Nanjian County, Lingbaoshan National Forest Park, 24°78'26"N, 100°51'30"E, altitude 2500 m a.s.l., on fallen angiosperm branch, leg. C.L. Zhao, 9 January 2019, CLZhao 10118, and CLZhao 10181 (SWFC).

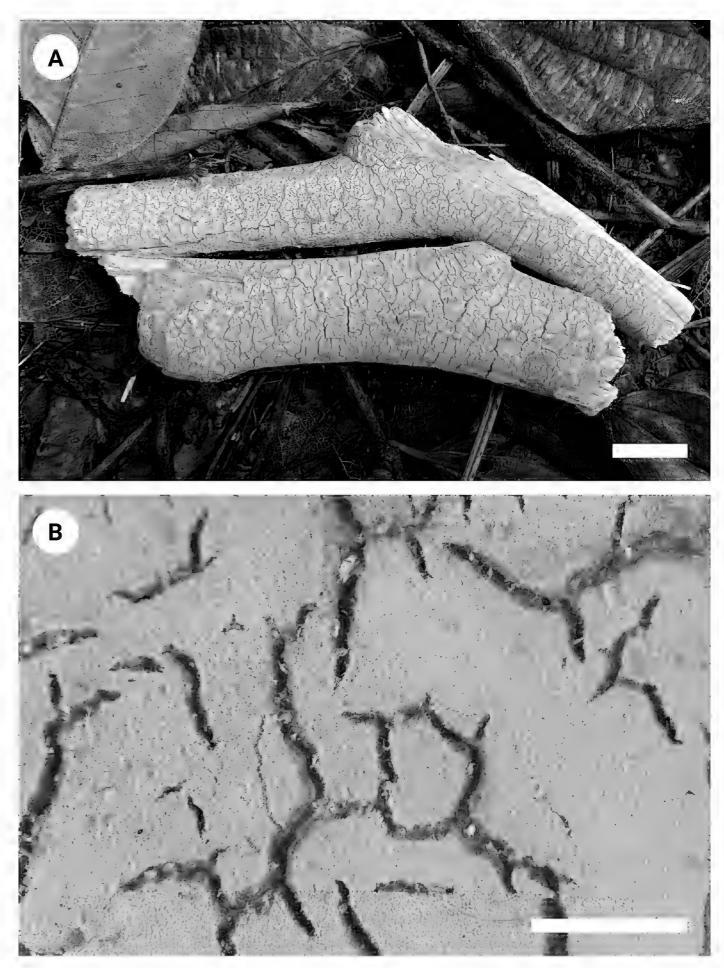


Figure 3. Basidiomata of Vararia fissurata (holotype). Scale bars: 1 cm (A); 1 mm (B).

Vararia isabellina Y.L. Deng & C.L. Zhao, sp. nov.

MycoBank No: MB851798

Figs 5, 6

Holotype. China. Yunnan Province, Lincang, Fengqing County, 24°67′18″N, 100°19′67″E, altitude 1660 m a.s.l., on the fallen angiosperm branch, leg. C.L. Zhao, 20 July 2022, CLZhao 22852 (SWFC).

Etymology. *Isabellina* (Lat.): referring to the isabelline to yellowish-brown basidiomata.

Description. Basidiomata annual, membranous, soft, and adnate, without odor or taste when fresh, up to 90 mm long, 10 mm wide, and $50-90 \mu m$ thick. Hymenial surface smooth, cream to isabelline when fresh, isabelline to slightly brown when dry. Sterile margin thinning out, cream to isabelline, and up to 1 mm wide.

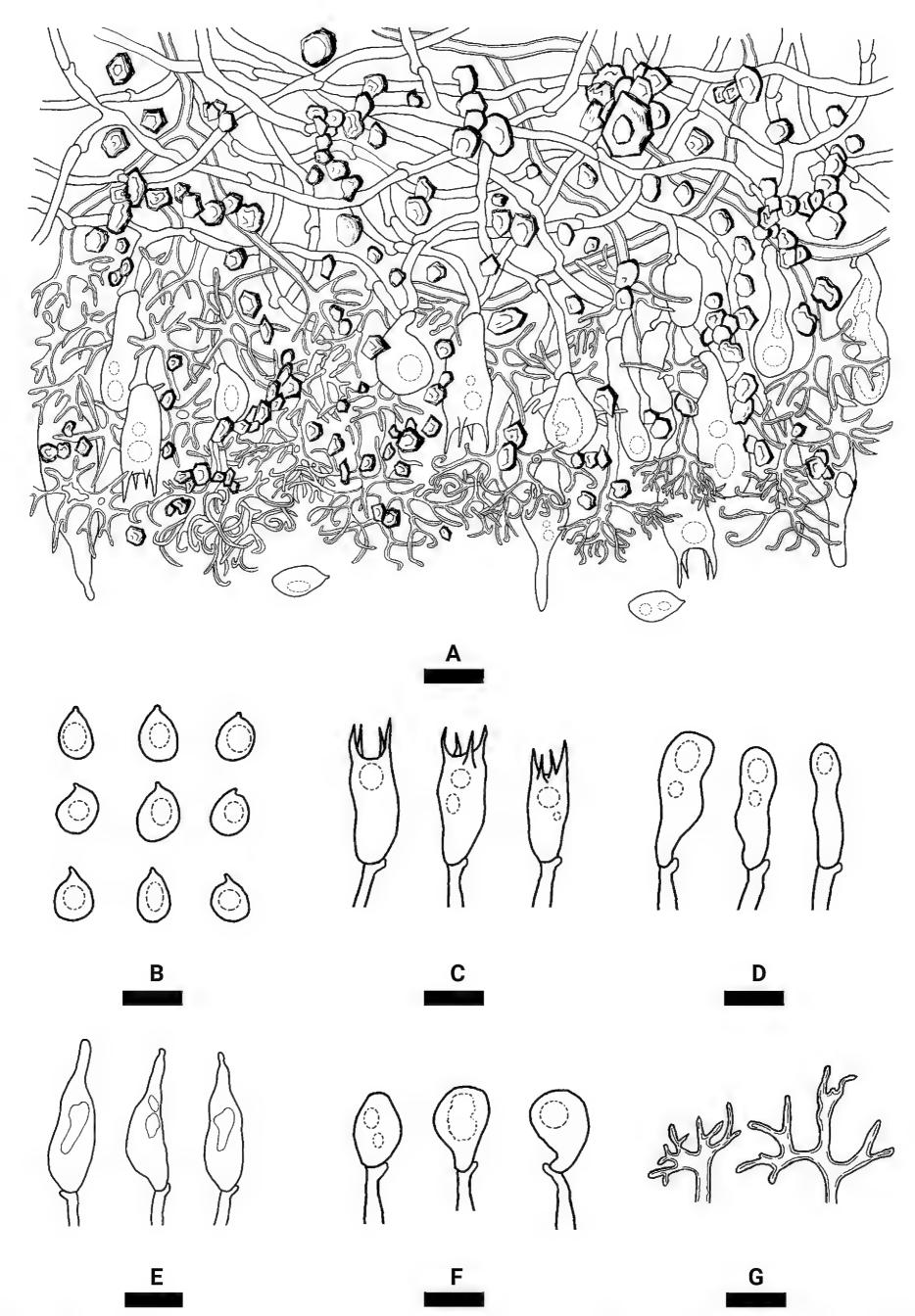


Figure 4. Microscopic structures of *Vararia fissurata* (holotype) **A** a section of hymenium **B** basidiospores **C** basidia **D** basidioles **E** gloeocystidia subulate **F** gloeocystidia subglobose **G** dichohyphae. Scale bars: $10 \, \mu m \, (A-G)$.

Hyphal system dimitic, generative hyphae bearing simple-septa, colorless, thin to slightly thick-walled, frequently branched, 2.5–4 μ m in diameter, IKI–, CB–, tissues unchanged in KOH. Dichohyphae predominant, yellowish, distinctly thick-walled, dichotomously to irregularly branched with main branches up to 4 μ m in diameter and with acute tips, moderately dextrinoid in Melzer's reagent, CB–, tissues unchanged in KOH; dichohyphae in hymenium similar to those in subiculum but more branched, with more narrow and shorter branches, with slightly curved tips and stronger.

Gloeocystidia spindle to subcylindrical, smooth, colorless, thin-walled, usually containing refractive materials, $38-47\times8-13~\mu m$. Basidia subcylindrical, slightly constricted at the neck, with four sterigmata and a basal simple septum connection, $33-39\times7-9~\mu m$; basidioles dominant, in shape similar to basidia, but slightly smaller.

Basidiospores sub-fusiform to navicular, colorless, smooth, with numerous oil-drops, thin-walled, IKI-, CB-, 9-13 \times 5-8 μ m, L = 11.66 μ m, W = 6.69 μ m, Q = 1.68-1.78 (n = 60/2).

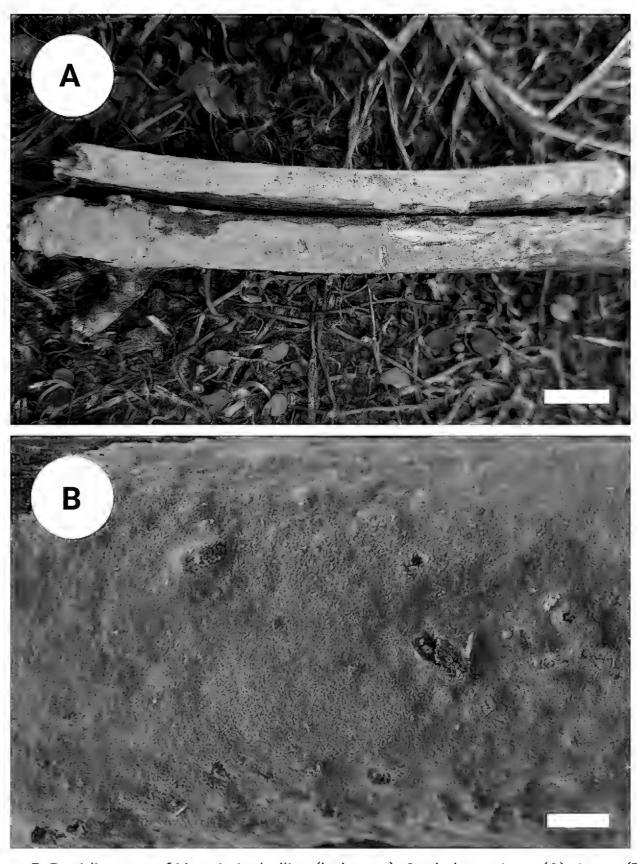


Figure 5. Basidiomata of Vararia isabellina (holotype). Scale bars: 1 cm (A); 1 mm (B).

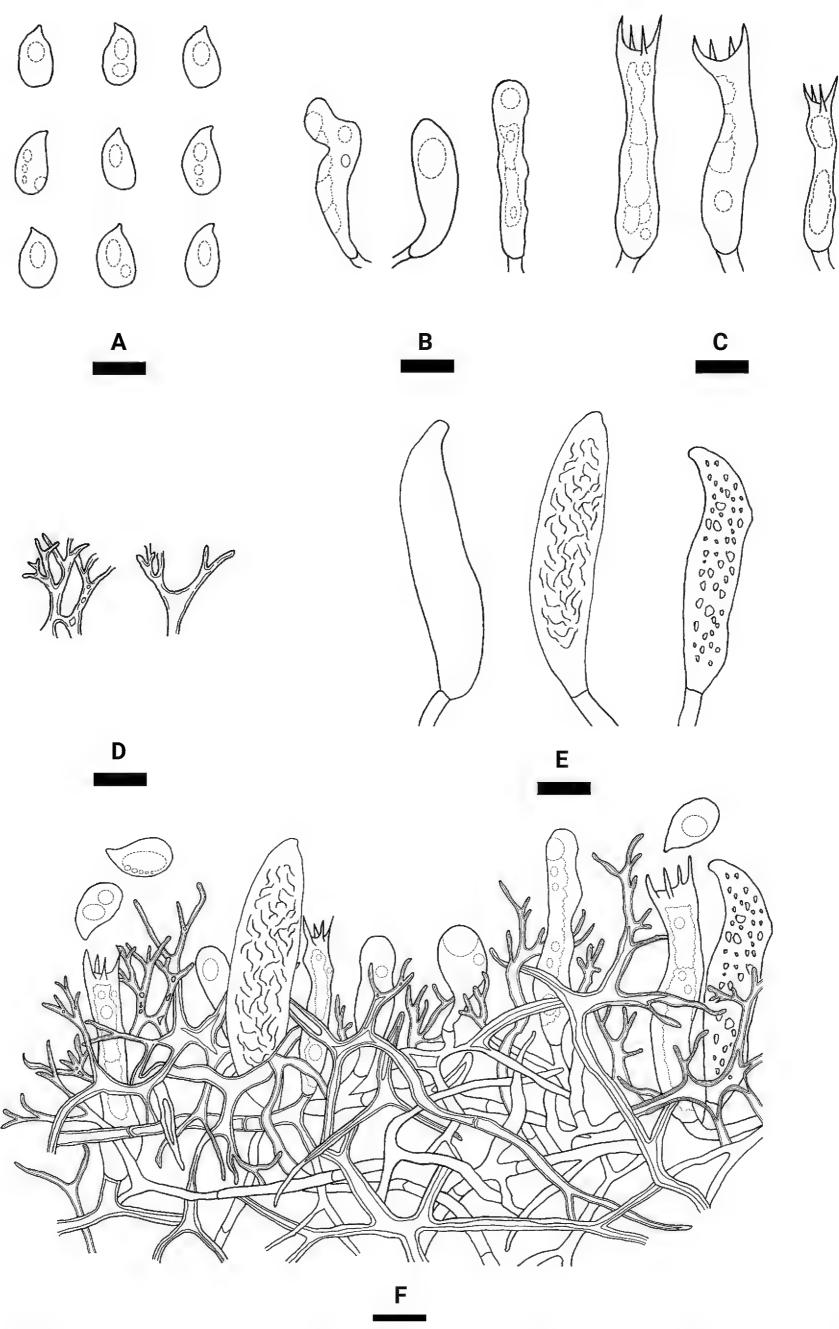


Figure 6. Microscopic structures of *Vararia isabellina* (holotype) **A** basidiospores **B** basidioles **C** basidia **D** dichohyphae **E** gloeocystidia **F** a section of hymenium. Scale bars: 10 μm (**A**–**F**).

Additional specimen examined (paratype). CHINA. Yunnan Province, Lincang, Fengqing County, 24°67′18″N, 100°19′67″E, altitude 1660 m a.s.l., on the fallen angiosperm branch, leg. C.L. Zhao, 20 July 2022, CLZhao 22887 (SWFC).

Vararia lincangensis Y.L. Deng & C.L. Zhao, sp. nov.

MycoBank No: MB851794

Figs 7, 8

Holotype. China. Yunnan Province, Lincang, Fengqing County, Yaojie Township, Xingyuan Village, 24°61'44"N, 100°17'21"E, altitude 1660 m a.s.l., on the fallen angiosperm branch, leg. C.L. Zhao, 20 July 2022, CLZhao 22791 (SWFC).

Etymology. *Lincangensis* (Lat.): referring to the locality (Lincang) of the type specimen.

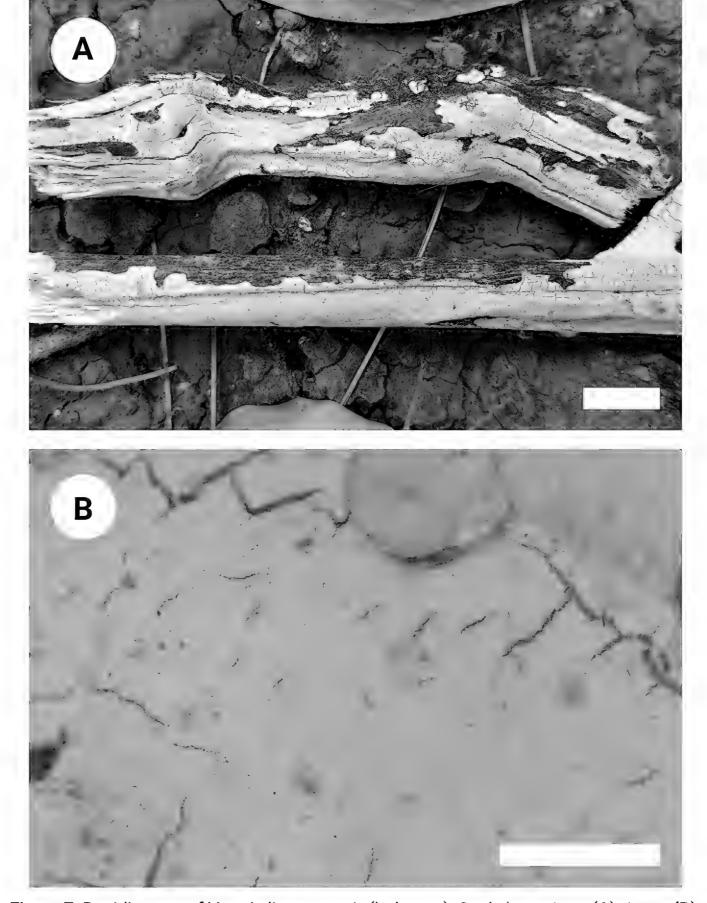


Figure 7. Basidiomata of Vararia lincangensis (holotype). Scale bars: 1 cm (A); 1 mm (B).

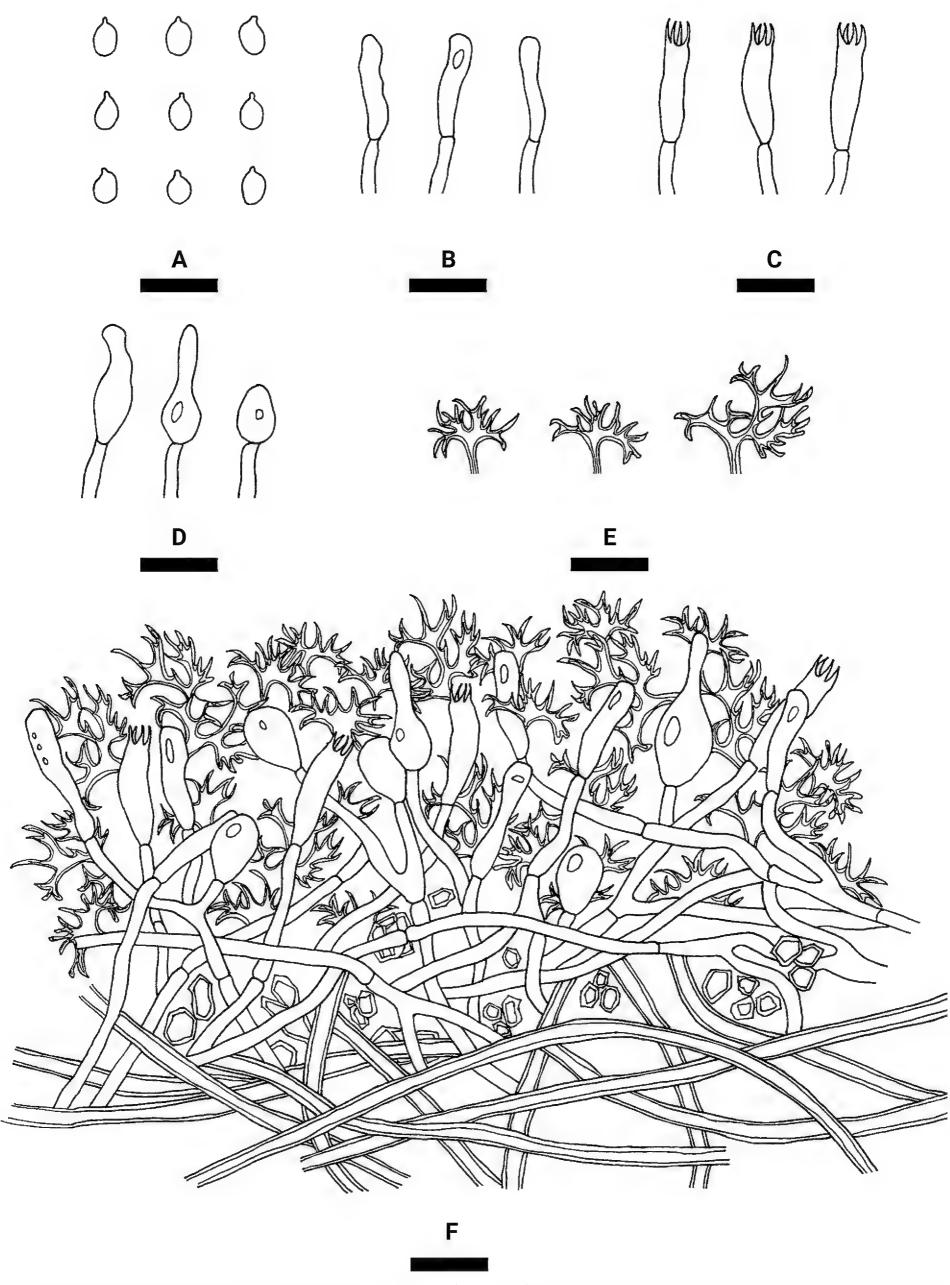


Figure 8. Microscopic structures of *Vararia lincangensis* (holotype) **A** basidiospores **B** basidioles **C** basidia **D** gloeocystidia **E** dichohyphae **F** a section of hymenium. Scale bars: $10 \mu m$ (**A–F**).

Description. Basidiomata annual, resupinate, membranous, soft and adnate, without odor or taste when fresh, up to 90 mm long, 20 mm wide, and 70–150 μm thick. Hymenial surface smooth, white to cream when fresh, cream upon drying, cracking with age. Sterile margin distinct, narrow, whitish, attached, and up to 1 mm wide.

Hyphal system dimitic, generative hyphae bearing simple-septa, rarely branched, colorless, thin-walled, 2–3 μ m in diameter, IKI–, CB–, tissues unchanged in KOH; subhymenial hyphae densely covered by some crystals. Dichohyphae predominate, white to cream, capillary, thick-walled, frequently branched, dichotomously to irregularly branched with main branches and acute tips, 1–1.5 μ m diameter, weakly to moderately dextrinoid in Melzer's reagent, CB–, tissues unchanged in KOH, subiculum composed of colorless. Skeletal hyphae colorless, thick-walled, 2–3 μ m in diameter, IKI–, CB–, tissues unchanged in KOH.

Gloeocystidia subglobose, and clavate to fusiform, usually containing refractive materials, colorless, smooth, thin-walled, $6.5-16\times3-5$ µm. Basidia clavate, with four sterigmata and a basal simple septum, thin-walled, smooth, $11-17.5\times2-4$ µm; basidioles in shape similar to basidia, but slightly smaller.

Basidiospores ellipsoid, colorless, thin-walled, smooth, occasionally acyanophilous, CB-, (3-)3.5-5.5(-6) × (2-)2.5-4 μ m, L = 4.18 μ m, W = 3.11 μ m, Q = 1.33-1.36 (n = 60/2).

Additional specimen examined (paratype). CHINA. Yunnan Province, Lincang, Fengqing County, Yaojie Township, Xingyuan Village, 24°61'44"N, 100°17'21"E, altitude 1660 m a.s.l., on the fallen angiosperm branch, leg. C.L. Zhao, 20 July 2022, CLZhao 22799 (SWFC).

Vararia punctata Y.L. Deng & C.L. Zhao, sp. nov.

MycoBank No: MB851795

Figs 9, 10

Holotype. China. Yunnan Province, Dali, Weishan Country, Qinghua Town, Green Peacock Nature Reserve, 25°23'35"N, 100°31'39"E, altitude 1500 m a.s.l., on the fallen branch of angiosperm, leg. C.L. Zhao, 18 July 2022, CLZhao 22439 (SWFC).

Etymology. Punctata (Lat.): referring to the species having cushion-shaped basidioma.

Description. Basidiomata annual, membranous, soft, adnate, without odor or taste when fresh, up to 50 mm long, 15 mm wide, and $90-150~\mu m$ thick. Hymenial surface smooth, and white to cream when fresh, cream when dry. Sterile margin thin, distinct, narrow, whitish, attached, and up to 1 mm.

Hyphal system dimitic, generative hyphae bearing simple-septa, color-less, thin to slightly thick-walled, rarely branched, interwoven, 2–3 μm in diameter, IKI–, CB–, tissues unchanged in KOH. Dichohyphae predominate, white to cream, capillary, frequently branched, thick-walled, 1 μm in diameter, dichotomously to irregularly branched with main branches and acute tips, weakly to moderately dextrinoid in Melzer's reagent, CB–, tissues unchanged in KOH. Skeletal hyphae colorless, thick-walled, 2–3 μm in diame-

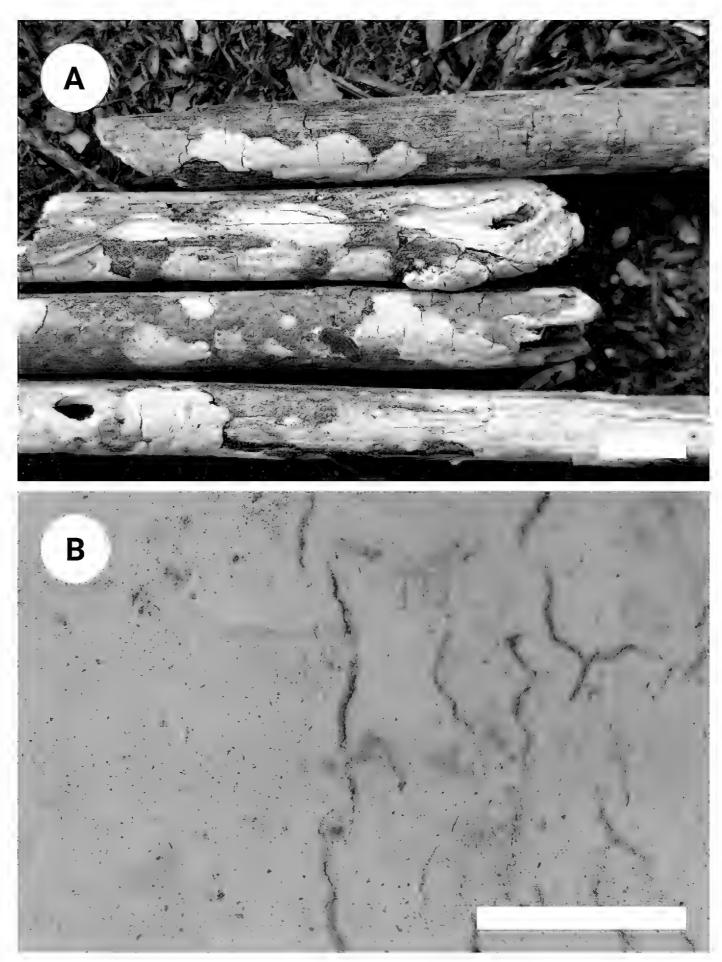


Figure 9. Basidiomata of Vararia punctata (holotype). Scale bars: 1 cm (A); 1 mm (B).

ter, IKI-, CB-, tissues unchanged in KOH; subhymenial hyphae densely covered by bulk crystals.

Gloeocystidia clavate to cylindrical, usually containing oil droplets, colorless, smooth, thick-walled, and $12-21\times5-9$ µm. Basidia subcylindrical, with four sterigmata and a basal simple septum, $11-25\times4-7$ µm; basidioles in shape similar to basidia, but slightly smaller.

Basidiospores ellipsoid, colorless, thin-walled, smooth, with oil drops, amyloid, CB-, 6-10 × 4-6(-6.5) μ m, L = 7.81 μ m, W = 5.1 μ m, Q = 1.51-1.56 (n = 120/4).

Additional specimen examined (paratype). CHINA. Yunnan Province, Dali, Weishan Country, Qinghua Town, Green Peacock Nature Reserve, 25°23'35"N, 100°31'39"E, altitude 1500 m a.s.l., on the fallen branch of angiosperm, leg. C.L. Zhao, 18 July 2022, CLZhao 22423 (SWFC).

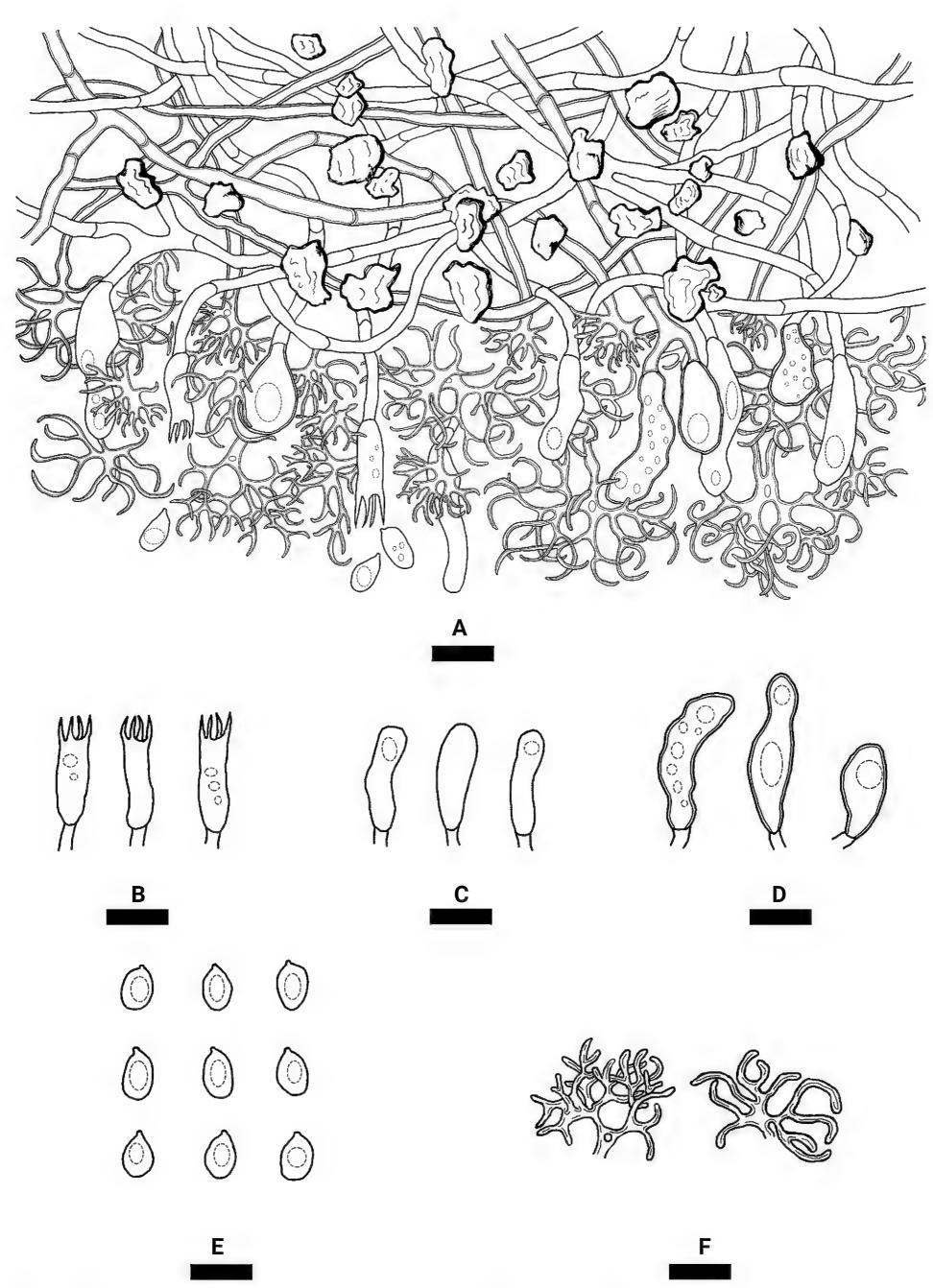


Figure 10. Microscopic structures of *Vararia punctata* (holotype) **A** a section of hymenium **B** basidia **C** basidioles **D** gloeocystidia **E** basidiospores **F** dichohyphae. Scale bars: $10 \mu m (A-F)$.

Vararia sinensis Y.L. Deng & C.L. Zhao, sp. nov.

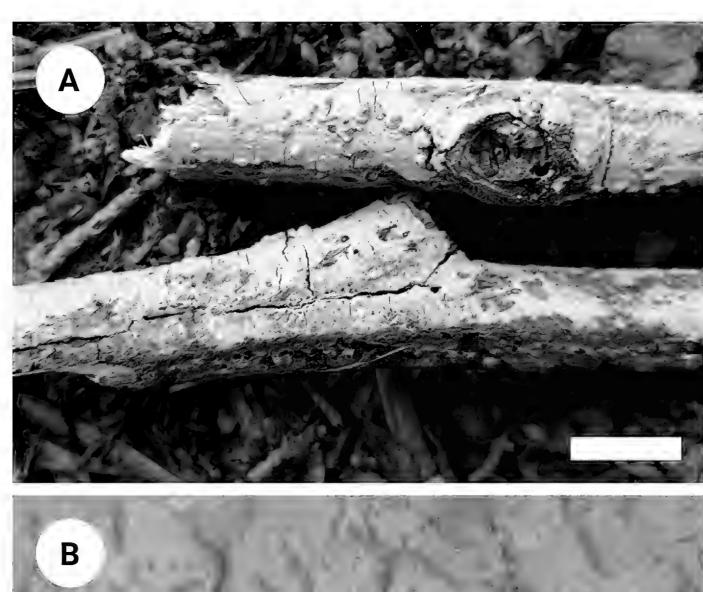
MycoBank No: MB851796

Figs 11, 12

Holotype. China. Yunnan Province, Lincang, Yun County, Dumu Village, 24°39'79"N, 100°18'17"E, altitude 1960 m a.s.l., on the fallen angiosperm branch, leg. C.L. Zhao, 20 October 2022, CLZhao 25160 (SWFC).

Etymology. Sinensis (Lat.): referring to the locality (China) of the type specimen.

Description. Basidiomata annual, membranous, and adnate, up to 70 mm long, 35 mm wide, and $80-160~\mu m$ thick. Hymenial surface smooth, white to slightly pink when fresh, pink upon drying. Sterile margin thinning out, narrow, whitish, attached, and up to 1 mm.



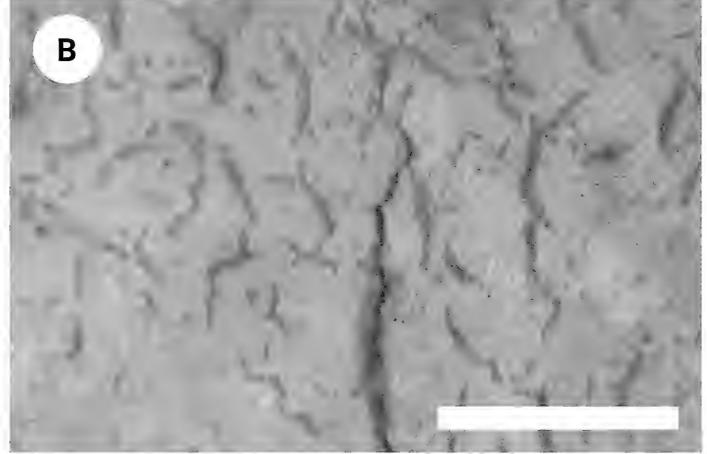


Figure 11. Basidiomata of Vararia sinensis (holotype). Scale bars: 1 cm (A); 1 mm (B).

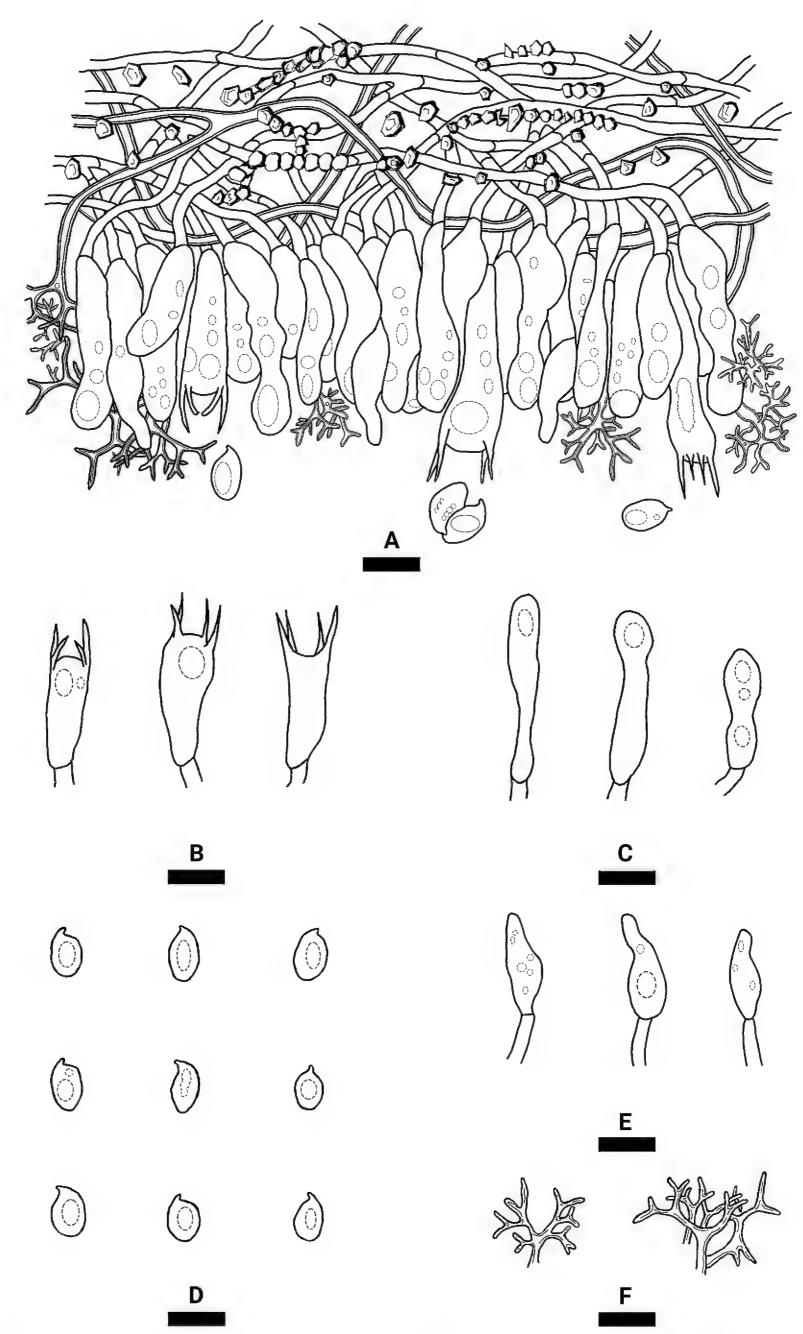


Figure 12. Microscopic structures of *Vararia sinensis* (holotype) **A** basidiospores **B** basidioles **C** basidia **D** dichohyphae **E** gloeocystidia subulate **F** a section of hymenium. Scale bars: $10 \mu m (A-F)$.

Hyphal system dimitic, generative hyphae bearing simple-septa, colorless, thin-walled, branched, 2.5–3 μ m diameter, IKI–, CB–, tissues unchanged in KOH. Dichohyphae predominant, yellowish, thick-walled, dichotomously to irregularly branched with main branches up to 1.4 μ m in diameter and with acute tips, moderately dextrinoid in Melzer's reagent, CB–, tissues unchanged in KOH, dichohyphae in hymenium similar to those in subiculum but more branched, with more narrow and shorter branches, with slightly curved tips and stronger, subhymenial hyphae densely covered by crystals. Skeletal hyphae rarely branched, interwoven, colorless, thick-walled, 2–3 μ m in diameter, IKI–, CB–, tissues unchanged in KOH.

Gloeocystidia subulate, smooth, colorless, thin-walled, filled with refractive oil-like matter, $17-35\times6-7~\mu m$. Basidia clavate, with four sterigmata and a basal simple septum connection, $25-35\times6-7~\mu m$; basidioles dominant, in shape similar to basidia, but slightly smaller.

Basidiospores sub-fusiform to navicular, with a beaklike extension, colorless, smooth, with numerous oil-drops, thin-walled, IKI-, CB-, 6-11 \times 4-6 μ m, L = 8.21 μ m, W = 4.88 μ m, Q = 1.66-1.71 (n = 60/2).

Additional specimen examined (paratype). CHINA. Yunnan Province, Lincang, Yun County, Dumu Village. GPS coordinates: 24°39'79"N, 100°18'17"E, altitude 1960 m a.s.l., on the fallen angiosperm branch, leg. C.L. Zhao, 20 October 2022, CLZhao 25161 (SWFC).

Vararia yaoshanensis Y.L. Deng & C.L. Zhao, sp. nov.

MycoBank No: MB851797

Figs 13, 14

Holotype. China. Yunnan Province, Zhaotong, Qiaojia County, Yao Shan National Nature Reserve, 26°89'62"N, 102°95'04"E, altitude 2500 m a.s.l., on fallen branch of angiosperm, 21 August 2020, CLZhao 20693 (SWFC).

Etymology. Yaoshanensis (Lat.): referring to the provenance (Yaoshan) of the type specimen.

Description. Basidiomata annual, membranous, adnate, without odor or taste when fresh, up to 8 cm long, 4 cm wide, 80–120 µm thick. Hymenial surface smooth, cream to cinnamon-buff when fresh, pinkish buff to cinnamon-buff upon drying, cracking with age. Sterile margin thin, pinkish buff, up to 1 mm.

Hyphal system dimitic, generative hyphae bearing simple-septa, rarely branched, slightly thick-walled, 2–4 μm in diameter, IKI–, CB–, tissues unchanged in KOH. Dichohyphae, predominant, capillary, frequently branched, distinctly thick-walled, 1.6 μm diameter, dichotomously to irregularly branched with main branches and acute tips, weakly to moderately dextrinoid in Melzer's reagent, CB–, tissues unchanged in KOH.

Gloeocystidia with two types, (1) Gloeocystidia fusiform, colorless, thick-walled, smooth, tapered or gradually elongated apex, $28.5-50\times6-12.5~\mu m$; (2) Gloeocystidia subglobose, usually containing refractive materials, colorless, thick-walled, smooth, $11-27\times7-11~\mu m$. Basidia are subclavate to subcylindrical, thin-walled, with four sterigmatas and a basal simple septum, $23-46\times5-8~\mu m$; basidioles dominant, in shape similar to basidia, but slightly smaller.

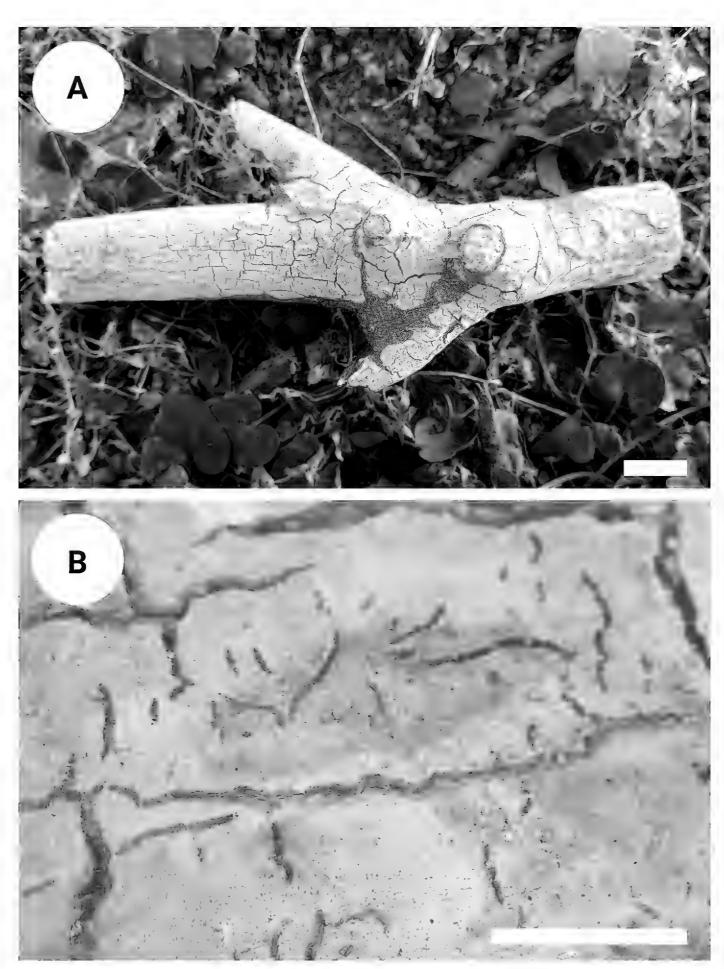


Figure 13. Basidiomata of Vararia yaoshanensis (holotype). Scale bars: 1 cm (A); 1 mm (B).

Basidiospores ellipsoid, colorless, slightly thick-walled, smooth, amyloid, CB-, $(7.5-)7.6-10.8(-10.9)\times(5.3-)5.7-7.8(-7.9)$ µm, L = 9.52 µm, W = 6.61 µm, Q = 1.4-1.5 (n = 210/7).

Additional specimens examined (paratypes). CHINA. Yunnan Province, Zhaotong, Qiaojia County, Yao Shan National Nature Reserve, 26°89'62"N, 102°95'04"E, altitude 2500 m a.s.l., on fallen branch of angiosperm, 21 August 2020, CLZhao 20669, CLZhao 20677, CLZhao 20697, CLZhao 20709, CLZhao 20713, CLZhao 20717 and CLZhao 20724 (SWFC), 22 August 2020, CLZhao 20528, CLZhao 20531, CLZhao 20565, CLZhao 20605, CLZhao 20608, CLZhao 20617, CLZhao 20619, CLZhao 20624, CLZhao 20646 and CLZhao 20656 (SWFC).

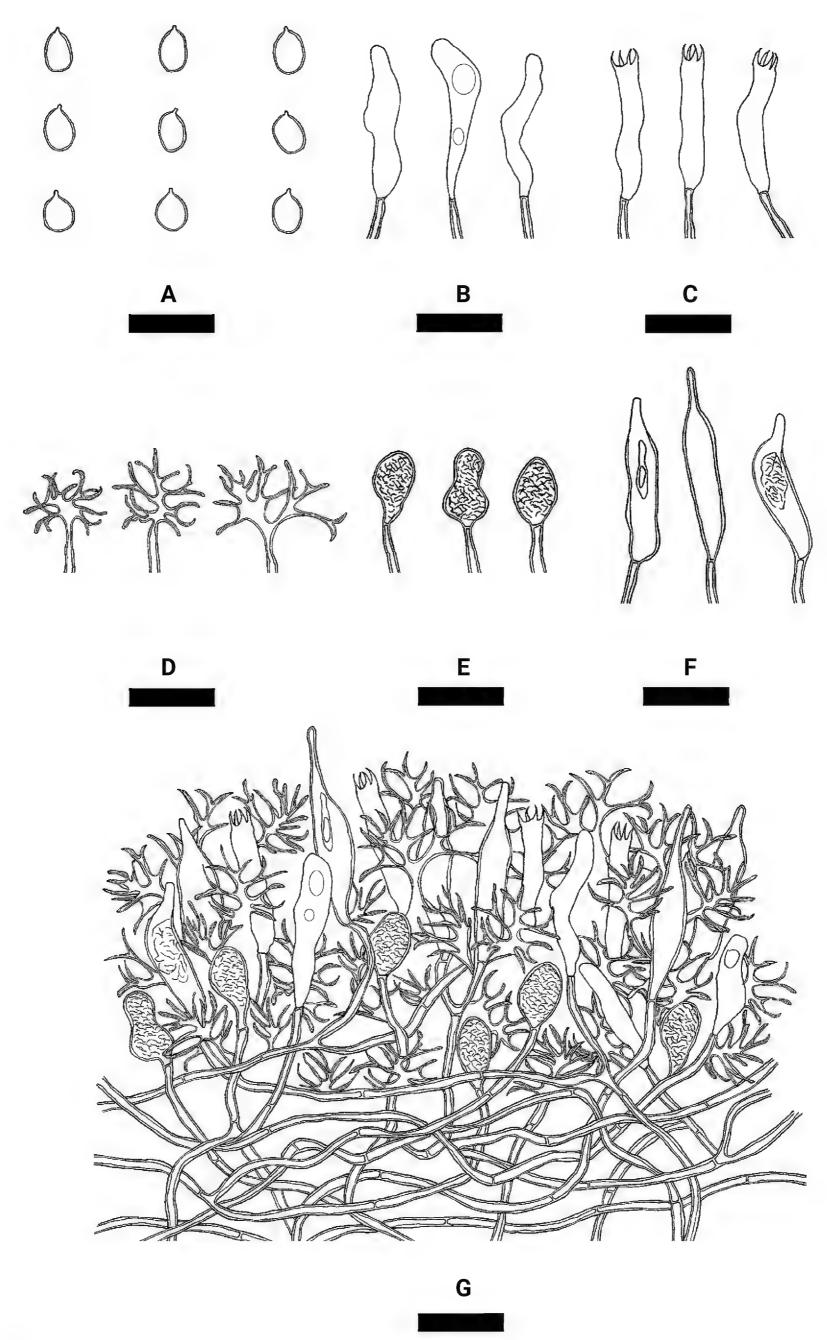


Figure 14. Microscopic structures of *Vararia yaoshanensis* (holotype) **A** basidiospores **B** basidia **C** basidioles **D** dichohyphae **E** gloeocystidia subglobose **F** gloeocystidia clavate to fusiform **G** a section of hymenium. Scale bars: $10 \mu m (A-G)$.

Discussion

Many recently described wood-inhabiting fungal taxa have been reported worldwide, including in the genera *Vararia* (Larsson 2007; Bernicchia and Gorjón 2010; Duhem and Buyck 2012; Sanyal et al. 2012; Nakasone 2015; Liu and He 2016; Leal-Dutra et al. 2018; Liu 2019; Dai et al. 2021; Zou et al. 2022; Deng and Zhao 2023; Li et al. 2023). Prior to this study, the following eleven *Vararia* species were reported from China, *V. amphithallica* Boidin, Lanq. & Gilles, *V. bispora* S.L. Liu & S.H. He, *V. breviphysa*, *V. cinnamomea* Boidin, Lanq. & Gilles, *V. daweishanensis*, *V. fragilis*, *V. investiens*, *V. montana* S.L. Liu & S.H. He, *V. racemosa* (Burt.) D.P. Rogers & H.S. Jacks., *V. sphaericospora* Gilb. and *V. yunnanensis* Y.L. Deng & C.L. Zhao (Dai 2011; Liu 2019; Dai et al. 2021; Zou et al. 2022; Deng and Zhao 2023). The present study (Figs 1, 2) reports six new species in *Vararia*, based on a combination of morphological features and molecular evidences.

Phylogenetically, based on the multiple loci in Scytinostroma s.s., nine genera, Asterostroma, Baltazaria, Dichostereum, Gloiothele, Lachnocladium, Michenera, Peniophora, Vesiculomyces and Vararia were divided in the family Peniophoraceae (Larsson and Larsson 2003, 2004; Larsson 2007; Leal-Dutra et al. 2018; Liu and He 2018; Zou et al. 2022; Li et al. 2023). In the present study, based on the ITS+LSU data (Fig. 1), Vararia was grouped with Asterostroma, Baltazaria, Dichostereum, and Peniophora, in which six new species were grouped into the genus Vararia. From the phylogram inferred from the ITS+LSU data (Fig. 1), the four new species V. fissurata, V. punctata, V. isabellina and V. sinensis were retrieved as a sister to V. ellipsospora, V. ambigua, V. investiens and V. breviphysa, respectively. Furthermore, the two new species Vararia lincangensis and V. yaoshanensis formed a monophyletic lineage respectively, and then V. yaoshanensis was clustered with V. ellipsospora and V. tropica. The species V. lincangensis was grouped closely with V. ambigua, V. gallica and V. punctata. However, morphologically, V. investiens can be delimited from V. isabellina by having the resupinate basidiomata with the yellowish cream to ochraceous hymenial surface, thin-walled, clamped generative hyphae, longer fusiform gloeocystidia ($40-80 \times 4-8 \mu m$), longer basidia ($30-50 \times 4-5 \mu m$), and smaller fusoid basidiospores measuring as 8-12 × 3-3.5 µm (Boidin and Languetin 1975). The taxon *V. ellipsospora* is different from *V. yaoshanensis* by having the smaller cylindrical basidia ($24-30 \times 5-6 \mu m$), longer gloeocystidia $(40-56 \times 8-10 \,\mu\text{m})$, and narrower basidiospores $(8-12 \times 5.5-6.5 \,\mu\text{m}$; Cunningham 1955), and V. tropica is distinguished from V. yaoshanensis by its smaller subcylindrical gloeocystides (20–42 \times 6.5–10 μ m), and larger basidia (35–50 \times 7–8.5 µm; Welden 1965). In addition, V. ambigua differs from V. lineangensis by having both larger gloeocystidia (15–32 \times 3.5–7 μ m), and basidiospores measuring as $6-7.3 \times 3.4-5 \,\mu m$ (Boidin et al. 1980); V. gallica is different from V. lincangensis by its larger fusiform gloeocystidia (15-36 × 3.5-6.5 μm) and basidiospores (9-12 \times 3.5-5 µm; Boidin and Languetin 1975; Grosse-Brauckmann and Kummer 2004).

Based on ITS topology (Fig. 2), the present study highlighted that *V. fissurata* was found to be the sister to *V. ellipsospora* with strong supports, and morphologically *V. ellipsospora* is different from *V. fissurata* by the fimbriate basidiomata, thick-walled generative hyphae, larger flexuous-cylindrical gloeocystid-

ia $(40-56 \times 8-10 \mu m)$, longer basidia $(24-30 \times 5-6 \mu m)$, and longer oblong ellipsoid basidiospores (8–12 \times 5.5–6.5 μ m; Cunningham 1955). In addition, V. lincangensis was clustered with V. fragilis, but morphologically V. fragilis is distinguished from V. lincangensis by the brittle basidiomata, with a buff to ochraceous hymenial surface and elliptical to ovoid gloeocystidia, both larger subulate gloeocystidia (16.5-27 \times 4-7 μ m) and subcylindrical basidia (13- $23.5 \times 3-4.5 \mu m$; Zou et al. 2022). Furthermore, V. punctata was retrieved as a sister to V. ambigua, but morphologically V. ambigua differs from V. punctata by its cream to buff hymenophore, and larger fusiform gloeocystidia measuring as 15-32 × 3.5-7 µm (Boidin et al. 1980). Further, V. isabellina formed a monophyletic lineage and then was grouped closely with V. daweishanensis and V. gracilispora Boidin & Lanq. However, morphologically V. daweishanensis is distinguishable from V. isabellina by its pale yellowish hymenial surface, clamped generative hyphae, and smaller gloeocystidia (9-23 \times 7-10.5 μ m), longer basidia measuring as 26-46 × 5-8 µm, narrower allantoid basidiospores (9-13 × 3.5-5 μm; Zou et al. 2022). Moreover, V. sinensis was grouped with five taxa: V. breviphysa, V. pirispora, V. fusispora, V. abortiphysa, and V. insolita, however, morphologically, V. breviphysa is distinguishable from V. sinensis by having light yellow to light brown basidiomata, larger subcylindrical gloeocystides (50-65 \times 6-8.5 μ m), larger basidia (30-38 \times 5.5-7 μ m), and longer fusiform basidiospores (16–20 × 4–5 μ m, Boidin and Lanquetin 1975; Liu et al. 2019); the species V. pirispora is distinct from V. sinensis by its larger subcylindrical gloeocystides ($40-65 \times 6-8 \mu m$), longer basidia measuring as $36-52 \times 6-7 \mu m$, larger pyriform basidiospores (10-16.5 × 5-7 μm ; Boidin et al. 1987); V. fusispora can be delimited from V. sinensis by having larger cylindrical gloeocystidia (40-60 × 5-6 µm) and oval to fusiform gloeocystidia $(24-60 \times 6-12 \,\mu\text{m})$, subclavate basidia $(35-56 \times 6-9 \,\mu\text{m})$, and larger fusiform basidiospores measuring as $14-17 \times 4-6 \mu m$ (Cunningham 1955); V. abortiphysa is distinct from V. sinensis by its plagio and subcylindrical gloeocystides measuring as $25-45 \times 4.5-9 \mu m$, and longer cylindrical basidiospores (14-17 × 2.2–2.8 μm; Boidin and Lanquetin 1975); V. insolita is distinguishable from V. sinensis by having larger gloeocystidia measuring as 60-80 × 5-8 μm, longer subcylindrical basidia $(30-78 \times 5.5-6.5 \mu m)$, and longer subfusiform basidiospores (12–16 × 4.2–5.75 µm; Boidin and Lanquetin 1975). Then V. yaoshanensis was found to be the sister to V. gallica (Bourdot & Galzin) Boidin with strong supports. However, morphologically, V. gallica can be delimited from V. yaoshanensis by its thin-walled generative hyphae, smaller thin-walled fusiform gloeocystidia (15–36 \times 3.5–6.5 μ m), and thin-walled, narrower basidiospores measuring as $9-12 \times 3.5-5 \mu m$ (Boidin and Languetin 1975; Grosse-Brauckmann and Kummer 2004).

Based on our phylogenetic and morphological research results, 17 species have been reported from China, including newly described in the present study and other recently published papers in this country (Dai 2011; Liu and He 2016; Liu 2019; Dai et al. 2021; Zou et al. 2022; Deng and Zhao 2023). It seems that the species diversity of *Vararia* is rich in China. Although *Vararia* taxa are well studied in the present paper, the species diversity, taxonomy and phylogeny of *Vararia* and related genera are still unresolved. A comprehensive study on this issue is urgently needed.

A key to 17 species of Vararia s.l. in China

1	Generative hyphae with clamp connections2
_	Generative hyphae bearing simple-septa3
2	Basidia with 2 sterigmatas4
_	Basidia with 4 sterigmatas5
3	Present thick-walled skeletal hyphae6
_	Absent thick-walled skeletal hyphae
4	Subcylindrical to fusiform basidiospores measuring as (10.5–)12–17(–
	20) × 4.5–5.5(–6.5) μ m, slightly thick-walled, subglobose gloeocystidia
	$(15-30(-35) \times 6-8(-10) \mu m)$, and subcylindrical or gradually narrower
_	gloeocystidia $(25-40(-65) \times 4.5-6(-18) \mu m)$
_	Fusiform to cylindrical basidiospores measuring as $(16-)18-22(-14) \times 6-7.2(-8) \mu m$, thick-walled, ventricose, gloeocystidia with an apical papil-
	la $(20-40 \times 9-12 \mu\text{m})$
5	Thin to thick-walled generative hyphae, subcylindrical basidia (26–46 ×
J	5-8 μ m), allantoid basidiospores measuring as (8.5-) 9-13 (-14) \times
	3.5–5 μ m, and ellipsoid to ovoid to subcylindrical gloeocystidia (9–23 ×
	7–10.5 μm)
_	Thin-walled generative hyphae8
6	Thin to slightly thick-walled generative hyphae, thick-walled, clavate to
	cylindrical gloeocystidia (12-21 × 5-9 μm), subcylindrical basidia (11-
	$25 \times 4-7 \mu\text{m}$), and ellipsoid basidiospores (6-10 × 4-6(-6.5) μ m)
	V. punctata
_	Thin-walled generative hyphae, clavate basidia9
7	Slightly thick-walled generative hyphae10
_	Thin-walled generative hyphae11
8	Gloeocystidia two kinds
_	Gloeocystidia one kinds
9	Ellipsoid basidiospores measuring as $(3-)3.5-5.5(-6) \times (2-)2.5-4 \mu m$,
	subglobose, clavate to fusiform gloeocystidia (6.5–16 × 3–5 μm)
_	Subfusiform to navicular basidiospores (6–11 × 4–6 μ m), subulate gloeo-
_	cystidia $(17-35 \times 6-7 \mu m)$
10	Slightly thick-walled, ellipsoid basidiospores measuring as (7.5–)7.6–
10	$10.8(-10.9) \times (5.3-)5.7-7.8(-7.9) \mu m$, thick-walled, fusiform gloeocys-
	tidia (28.5–50 × 6–12.5 μ m), globose gloeocystidia (11–27 × 7–11 μ m),
	subclavate to subcylindrical basidia $(23-46 \times 5-8 \mu m)$
	V. yaoshanensis
_	Thin-walled basidiospores, subcylindrical basidia13
11	Slightly thick-walled, ellipsoid basidiospores measuring as (5.1-)5.9-
	$11.5(-11.8) \times (4.3-)4.7-8.6(-9)$ µm, cylindrical basidia (17.5-32 ×
	$5-9.5~\mu m$), thin- to slightly thick-walled, subcylindrical gloeocystidia
	(16.5–58.5 × 4–10 μ m), fusiform gloeocystidia (18.5–43.5 × 7–9 μ m), ta-
	pering gloeocystidia (27.5–42 × 5.5–9 μ m)
-	Thin-walled basidiospores14
12	Basidiospores < 5 µm in diameter
_	Basidiospores > 5 µm in diameter15

13	Broad ellipsoid to ellipsoid basidiospores measuring as $3.5-5.5(-6) \times 10^{-6}$
	2.5–3.5 μ m, elliptical to ovoid gloeocystidia (5.8–16 × 3.5–7 μ m), subu-
	late gloeocystidia (16.5–27 × 4–7 μ m)
_	Sub-fusiform to navicular basidiospores with numerous oil-drops measur-
	ing as 9–13 \times 5–8 μ m, spindle to subcylindrical gloeocystidia (38–47 \times
	8–13 μm)
14	Rose to orange subfusiform basidiospores measuring as (14-)16-19(-
	21.5) × 4.2–6 μ m, cylindrical basidia (30–53 × 6.5–7.5 μ m), thick-walled,
	subcylindrical Gloeocystides (50–65 × 6–7(–8.5) μ m)
_	Colorless basidiospores
15	Broadly ellipsoid basidiospores measuring as $(11-)12-16(-17) \times (7.5-$
)9.5–13(–14) μ m, clavate basidia (70–110 × 10–16 μ m), clavate gloeo-
	cystidia (50–100 × 4–9 μm)
_	Spherical basidiospores measuring as $8-10 \times 7.5-8.5 \mu m$, cylindrical to
	clavate basidia ($40-45 \times 6-7.5 \mu m$), subcylindrical to fusiform gloeocys-
	tides $(48-80(-105) \times 7-11(-14) \mu m)$
16	Subcylindrical to fusiform gloeocystides (26–40 \times 4.5–9 μ m), cylindrical
	basidiospores (6–8 × 2–3 μ m), cylindrical basidia (30–40 × 4–5 μ m)
_	Absent gloeocystides, oblong to subellipsoid basidiospores measuring as
	$9-13 \times 5-7.2 \mu m$, and subcylindrical basidia ($45-65 \times 8-10 \mu m$)
	V. cinnamomea

Additional information

Conflict of interest

The authors have declared that no competing interests exist.

Ethical statement

No ethical statement was reported.

Funding

The research was supported by the National Natural Science Foundation of China (Project Nos. 32170004, U2102220), and the High-level Talents Program of Yunnan Province (YNQR-QNRC-2018-111), and Forestry Innovation Programs of Southwest Forestry University (Grant No: LXXK-2023Z07).

Author contributions

Conceptualization: CZ. Data curation: YD, CZ. Formal analysis: CZ, YD, SJ. Funding acquisition: CZ. Investigation: YD, CZ, SJ. Methodology: SJ, YD, CZ. Project administration: CZ. Resources: CZ, YD. Software: YD, CZ. Supervision: YD, SJ, CZ. Validation: CZ. Visualization: CZ. Writing – original draft: YD, CZ, SJ. Writing – review and editing: YD, CZ.

Author ORCIDs

Yinglian Deng https://orcid.org/0000-0002-8220-508X Sana Jabeen https://orcid.org/0000-0001-8839-7716 Changlin Zhao https://orcid.org/0000-0002-8668-1075

Data availability

All of the data that support the findings of this study are available in the main text.

References

- Bernicchia A, Gorjón SP (2010) Fungi Europaei 12: *Corticiaceae* s.l. Edizioni Candusso, Alassio, Italy.
- Boidin J (1967) Basidiomycètes Lachnocladiaceae résupinés de la Republique Centrafricaine. Cahiers de La Maboké 5: 23–35.
- Boidin J (1980) Application du concept biologique del'espèce aux Basidiomycètes. Le genre *Vararia* section *Vararia* au Gabon. Cryptogamie. Mycologie 1: 265–384.
- Boidin J, Gilles G (1999) Contribution à la connaissance du genre *Vararia* (Basidiomycotina). Bulletin de la Societe Mycologique de France 115: 115–139.
- Boidin J, Lanquetin P (1975) *Vararia* subgenus *Vararia* (Basidiomycetes, Lachnocladiaceae): Étude spèciale des espèces d'Afrique intertropicale. Bulletin de la Société Mycologique de France 91: 457–513.
- Boidin J, Lanquetin P (1987) Le genre *Scytinostroma* Donk (Basidiomycetes, Lachnocladiaceae). Bibliotheca Mycologica 114: 1–130.
- Boidin J, Lanquetin P, Gilles G (1980) Application du concept biologique del'espèce aux Basidiomycètes. Le genre *Vararia* section *Vararia* au Gabon. Cryptogamie. Mycologie 1(4): 265–384.
- Boidin J, Gilles G, Lanquetin P (1987) Basidiomycètes Aphyllophorales de l'Île de la Réunion. IX Les genres *Dichostereum* Pilat et *Vararia* Karsten. Bulletin de la Société Mycologique de France 103(2): 119–135.
- Boidin J, Mugnier J, Canales R (1998) Taxonomie moleculaire des Aphyllophorales. Mycotaxon 66: 445–491.
- Cui BK, Li HJ, Ji X, Zhou JL, Song J, Si J, Yang ZL, Dai YC (2019) Species diversity, taxonomy and phylogeny of Polyporaceae (Basidiomycota) in China. Fungal Diversity 97(1): 137–392. https://doi.org/10.1007/s13225-019-00427-4
- Cunningham GH (1955) Thelephoraceae of New Zealand. Part IV. The genus *Vararia*. Transactions and Proceedings of the Royal Society of New Zealand 82: 973–985.
- Dai YC (2011) A revised checklist of corticioid and hydnoid fungi in China for 2010. Mycoscience 52(1): 69–79. https://doi.org/10.1007/S10267-010-0068-1
- Dai YC, Yang ZL, Cui BK, Wu G, Yuan HS, Zhou LW, He SH, Ge ZW, Wu F, Wei YL, Yuan Y, Si J (2021) Diversity and systematics of the important macrofungi in Chinese forests. Mycosystema 40: 770–805. https://doi.org/10.13346/j.mycosystema.210036
- Deng YL, Zhao CL (2023) The molecular phylogeny and morphology revealed a new wood-rotting fungus *Vararia yunnanensis* (Peniophoraceae, Russulales) in Yunnan Province, China. Phytotaxa 583: 039–049. https://doi.org/10.11646/phytotaxa.583.1.4
- Duhem B, Buyck B (2012) On two new tropical *Vararia* (Russulales, Basidiomycota) with extremely small, racemose dichohyphidia. Cryptogamie. Mycologie 33(4): 427–437. https://doi.org/10.7872/crym.v33.iss4.2012.427
- Felsenstein J (1985) Confidence intervals on phylogenetics: An approach using bootstrap. Evolution; International Journal of Organic Evolution 39(4): 783–791. https://doi.org/10.2307/2408678
- Gilbertson RL (1965) Some species of *Vararia* from temperate North America. Papers of the Michigan Academy of Science, Arts and Letters 50: 161–184.

- Grosse-Brauckman H, Kummer V (2004) Fünf bemerkenswerte funde corticioider Pilze aus Deutschland. Feddes Repertorium 115(1): 90–101. https://doi.org/10.1002/fedr.200311029
- Guan QX, Huang J, Huang J, Zhao CL (2023) Five new species of Schizoporaceae (Basidiomycota, Hymenochaetales) from East Asia. MycoKeys 96: 25–56. https://doi.org/10.3897/mycokeys.96.99327
- Hallenberg N (1985) The Lachnocladiaceae and Coniophoraceae of North Europe. Fungiflora.
- Hyde KD (2022) The numbers of fungi. Fungal Diversity 114(1): 1. https://doi. org/10.1007/s13225-022-00507-y
- Jiang N, Voglmayr H, Bian DR, Piao CG, Wnag SK, Li Y (2021) Morphology and phylogeny of *Gnomoniopsis* (Gnomoniaceae, Diaporthales) from fagaceae leaves in China. Journal of Fungi 7(10): 792. https://doi.org/10.3390/jof7100792
- Karasinski D (2010) Polish resupinate Russulales: The genus *Vararia*. Acta Mycologica 45(1): 45–56. https://doi.org/10.5586/am.2010.007
- Karnste PA (1898) Kritisk of versigt af Finlands Basidsvampar. Biology 3: 1-36.
- Katoh K, Rozewicki J, Yamada KD (2019) MAFFT online service: Multiple sequence alignment, interactive sequence choice and visualization. Briefings in Bioinformatics 20(4): 1160–1166. https://doi.org/10.1093/bib/bbx108
- Larsson KH (2007) Re-thinking the classification of corticioid fungi. Mycological Research 111(9): 1040–1063. https://doi.org/10.1016/j.mycres.2007.08.001
- Larsson A (2014) AliView: A fast and lightweight alignment viewer and editor for large data sets. Bioinformatics 30(22): 3276–3278. https://doi.org/10.1093/bioinformatics/btu531
- Larsson E, Larsson KH (2003) Phylogenetic relationships of russuloid basidiomycetes with emphasis on aphyllophoralean taxa. Mycologia 95(6): 1037–1065. https://doi.org/10.1080/15572536.2004.11833020
- Larsson KH, Larsson E, Kõljalg U (2004) High phylogenetic diversity among corticioid homobasidiomycetes. Mycological Research 108: 983–1002. https://doi.org/10.1017/S0953756204000851
- Leal-Dutra CA, Neves MA, Griffith GW, Reck MA, Clasen LA, Dentinger BTM (2018) Reclassification of *Parapterulicium* Corner (Pterulaceae, Agaricales), contributions to Lachnocladiaceae and Peniophoraceae (Russulales) and introduction of *Baltazaria* gen. nov. MycoKeys 37: 39–56. https://doi.org/10.3897/mycokeys.37.26303
- Li Y, Xu WQ, Liu SL, Yang N, He SH (2023) Species diversity and taxonomy of *Scytinostroma* sensu stricto (Russulales, Basidiomycota) with descriptions of four new species from China. MycoKeys 98: 133–152. https://doi.org/10.3897/mycokeys.98.105632
- Liu SL (2019) Taxonomy and phylogeny of *Vararia* and related genera in China. Ph.D. Thesis, Beijing Forestry University, Beijing, China.
- Liu SL, He SH (2016) The genus *Vararia* (Russulales, Basidiomycota) in China. Two new species and two new Chinese records. Nordic Journal of Botany 1756–1051. https://doi.org/10.1111/njb.01170
- Liu SL, He SH (2018) Taxonomy and phylogeny of *Dichostereum* (Russulales), with descriptions of three new species from southern China. MycoKeys 40: 111–126. https://doi.org/10.3897/mycokeys.40.28700
- Liu SL, Tian Y, Nie T, Thawthong A, Hyde KD, Xu LL, He SH (2017) Updates on East Asian *Asterostroma* (Russulales, Basidiomycota): New species and new records from Thailand and China. Mycological Progress 16(6): 667–676. https://doi.org/10.1007/s11557-017-1301-5

- Liu SL, Nakasone KK, He SH (2019) *Michenera incrustata* sp. nov. (Peniophoraceae, Russulales) from southern China. Nova Hedwigia 108(1-2): 197–206. https://doi.org/10.1127/nova_hedwigia/2018/0500
- Miller SL, Larsson E, Larsson KH, Verbeken A, Nuytinck J (2006) Perspectives in the new Russulales. Mycologia 98(6): 960–970. https://doi.org/10.1080/15572536.2006.11 832625
- Miller MA, Pfeiffer W, Schwartz T (2012) The CIPRES Science Gateway: enabling high-impact science for phylogenetics researchers with limited resources. Association for Computing Machinery 39: 1–8. https://doi.org/10.1145/2335755.2335836
- Nakasone KK (2015) Taxonomic studies in *Chrysoderma*, *Corneromyces*, *Dendro-physellum*, *Hyphoradulum*, and *Mycobonia*. Mycotaxon 130: 369–397. https://doi.org/10.5248/130.369
- Nylander JAA (2004) MrModeltest v.2. Program Distributed by the Author; Evolutionary Biology Centre, Uppsala University: Uppsala, Sweden.
- Petersen JH (1996) The danish mycological society's colour-chart. Foreningen til Svampekundskabens Fremme, Greve.
- Pouzar Z (1982) Taxonomic studies in resupinate fungi I. Czech Mycology 36: 141–145. Rehner SA, Samuels GJ (1994) Taxonomy and phylogeny of *Gliocladium* analysed from nuclear large subunit ribosomal DNA sequences. Mycological Research 98(6): 625–634. https://doi.org/10.1016/S0953-7562(09)80409-7
- Riebesehl J, Langer E (2017) *Hyphodontia* s.l. (Hymenochaetales, Basidiomycota): 35 new combinations and new keys to currently all 120 species. Mycological Progress 16(6): 637–666. https://doi.org/10.1007/s11557-017-1299-8
- Riebesehl J, Yurchenko E, Nakasone KK, Langer E (2019) Phylogenetic and morphological studies in *Xylodon* (Hymenochaetales, Basidiomycota) with the addition of four new species. MycoKeys 47: 97–137. https://doi.org/10.3897/mycokeys.47.31130
- Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Hohna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) MrBayes 3.2: Efficient Bayesian phylogenetic inference and model choice across a large model space. Systematic Biology 61(3): 539–542. https://doi.org/10.1093/sysbio/sys029
- Rosenthal LM, Larsson KH, Branco S, Chung JA, Glassman SI, Liao HL, Peay KG, Smith DP, Talbot JM, Taylor JW, Vellinga EC, Vilgalys R, Bruns TD (2017) Survey of corticioid fungi in North American pinaceous forests reveals hyperdiversity, underpopulated sequence databases, and species that are potentially ectomycorrhizal. Mycologia 109(1): 115–127. https://doi.org/10.1080/00275514.2017.1281677
- Sanyal SK, Dhingra GS, Singh AP (2012) *Vararia longicystidiata* sp. nov. (Agaricomycetes) from India. Mycotaxon 120(1): 357–360. https://doi.org/10.5248/120.357
- Stalpers JA (1996) The aphyllophoraceous fungi II. Keys to the species of the Hericiales. Studies in Mycology 40: 1–183.
- Vilgalys R, Hester M (1990) Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. Journal of Bacteriology 172(8): 4238–4246. https://doi.org/10.1128/jb.172.8.4238-4246.1990
- Vu D, Groenewald M, Vries M, Gehrmann T, Stielow B, Eberhardt U (2019) Large-scale generation and analysis of filamentous fungal DNA barcodes boosts coverage for kingdom fungi and reveals thresholds for fungal species and higher taxon delimitation. Studies in Mycology 92(1): 135–154. https://doi.org/10.1016/j.simy-co.2018.05.001
- Welden AL (1965) West Indian species of *Vararia* with notes on extralimital species. Mycologia 57(4): 502–520. https://doi.org/10.1080/00275514.1965.12018236

- White TJ, Bruns T, Lee S, Taylor J (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ (Eds) PCR protocols: A Guide to Methods And Applications. Academic Press, San Diego, 315–322. https://doi.org/10.1016/B978-0-12-372180-8.50042-1
- Wu F, Zhou LW, Vlasák J, Dai YC (2022) Global diversity and systematics of Hymeno-chaetaceae with poroid hymenophore. Fungal Diversity 113(1): 1–192. https://doi.org/10.1007/s13225-021-00496-4
- Yurchenko E, Wu SH (2016) A key to the species of *Hyphodontia* sensu lato. MycoKeys 12: 1–27. https://doi.org/10.3897/mycokeys.12.7568
- Yurchenko E, Riebesehl J, Langer E (2017) Clarification of *Lyomyces sambuci* complex with the descriptions of four new species. Mycological Progress 16(9): 865–876. https://doi.org/10.1007/s11557-017-1321-1
- Yurchenko E, Riebesehl J, Langer E (2020) *Fasciodontia* gen. nov. (Hymenochaetales, Basidiomycota) and the taxonomic status of *Deviodontia*. Mycological Progress 19(2): 171–184. https://doi.org/10.1007/s11557-019-01554-7
- Zhao CL, Wu ZQ (2017) *Ceriporiopsis kunmingensis* sp. nov. (Polyporales, Basidiomycota) evidenced by morphological characters and phylogenetic analysis. Mycological Progress 16(1): 93–100. https://doi.org/10.1007/s11557-016-1259-8
- Zhao YN, He SH, Nakasone KK, Wasantha KL, Chen CC, Liu SL, Ma HX, Huang MR (2021) Global phylogeny and taxonomy of the wood-decaying fungal genus *Phlebiopsis* (Polyporales, Basidiomycota). Frontiers in Microbiology 12: 622460. https://doi.org/10.3389/fmicb.2021.622460
- Zhao CL, Qu MH, Huang RX, Karunarathna SC (2023) Multi-gene phylogeny and taxonomy of the wood-rotting fungal genus *Phlebia* sensu lato (Polyporales, Basidiomycota). Journal of Fungi 9(3): 1–41. https://doi.org/10.3390/jof9030320
- Zou L, Zhang XL, Deng YL, Zhao CL (2022) Four new wood-inhabiting fungal species of Peniophoraceae (Russulales, Basidiomycota) from the Yunnan-Guizhou Plateau, China. Journal of Fungi 8(11): 1227. https://doi.org/10.3390/jof8111227